Total Synthesis and Biological Evaluation of Irciniastatin A (a.k.a. Psymberin) and Irciniastatin B

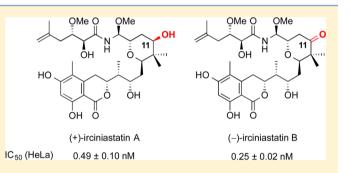
Shun-ichiro Uesugi,[†] Tsubasa Watanabe,[†] Takamichi Imaizumi,[†] Yu Ota,[‡] Keisuke Yoshida,[‡] Haruna Ebisu,[‡] Takumi Chinen,[‡] Yoko Nagumo,^{‡,§} Masatoshi Shibuya,[†] Naoki Kanoh,[†] Takeo Usui,^{*,‡,§} and Yoshiharu Iwabuchi^{*,†}

[†]Department of Organic Chemistry, Graduate School of Pharmaceutical Sciences, Tohoku University, Aobayama, Sendai 980-8578, Japan

[‡]Graduate School of Life and Environmental Sciences, University of Tsukuba, Tennodai, Tsukuba, Ibaraki 305-8572, Japan [§]Faculty of Life and Environmental Sciences, University of Tsukuba, Tennodai, Tsukuba, Ibaraki 305-8572, Japan

Supporting Information

ABSTRACT: Irciniastatin A (a.k.a. psymberin) and irciniastatin B are members of the pederin natural product family, which have potent antitumor activity and structural complexity. Herein, we describe a full account of our total synthesis of (+)-irciniastatin A and (-)-irciniastatin B. Our synthesis features the highly regioselective $Eu(OTf)_3$ -catalyzed, DTBMP-assisted epoxide ring opening reaction with MeOH, which enabled a concise synthesis of the C1–C6 fragment, extensive use of AZADO (2-azaadamantane *N*-oxyl) and its related nitroxyl radical/oxoammonium salt-catalyzed alcohol oxidation throughout the synthesis, and a late-stage assembly



of C1–C6, C8–C16, and C17–C25 fragments. In addition, for the synthesis of (-)-irciniastatin B, we achieved the C11selective control of the oxidation stage via regioselective deprotection and AZADO-catalyzed alcohol oxidation. The synthetic irciniastatins showed high levels of cytotoxic activity against mammalian cells. Furthermore, chemical footprinting experiments using synthetic compounds revealed that the binding site of irciniastatins is the E-site of the ribosome.

INTRODUCTION

In 2004, (+)-irciniastatin A (1) and (-)-irciniastatin B (2) were isolated by Pettit and co-workers¹ from the marine sponge *Ircinia ramosa*. Independently, psymberin was isolated by Crews and co-workers² from the marine sponge *Psammocinia* in the same year (Figure 1). These two reports revealed that these

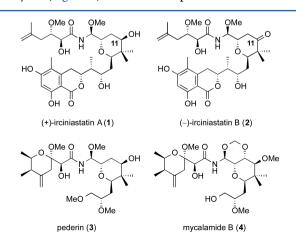


Figure 1. Pederin-type natural products.

natural products possessed a complex molecular architecture consisting of an unstable *N*,*O*-aminal moiety, a highly substituted 2,6-*trans*-tetrahydropyran skeleton, and either nine or eight stereogenic centers. At that time, Crews assumed that irciniastatin A and psymberin are identical; however, the stereochemistry of both compounds was not determined completely.

In 2005, the first total synthesis of (+)-irciniastatin A (1) was accomplished by De Brabander and co-workers,³ who established the absolute configuration of (+)-1 and demonstrated that (+)-irciniastatin A and psymberin possess identical structures. After this pioneering synthesis, 7 total syntheses⁴ including the synthesis by our laboratory,^{4d} 2 formal syntheses,⁵ and 10 synthetic studies⁶ have been reported.

Besides their fascinating structures, both (+)-irciniastatin A (1) and (-)-irciniastatin B (2) show strong inhibitory activity against a series of human cancer cell lines at the nanomolar level.¹ Interestingly, these two congeners possess different values of activity against several human cancer cell lines,¹ although these structures were considered to differ only in the oxidation level at C11. Subsequently, some analogues of 1 were

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synthesized and tested for their biological activities,^{4d,e,g,7} which gave highly potent derivatives and biological insights.⁸

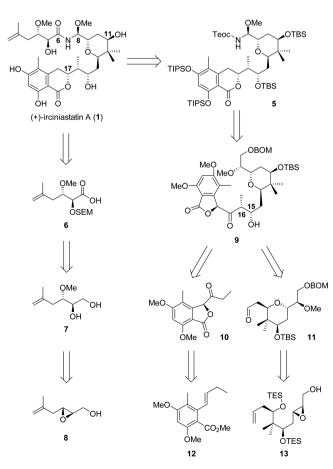
However, compared with those of (+)-irciniastatin A (1), the structural and biological details of (-)-irciniastatin B (2) have not been intensively investigated. In 2012, Smith and co-workers achieved the first total synthesis and determination of the absolute configuration of (-)-irciniastatin B (2),⁹ which is the only example of total synthesis.

To shed light on the impact of C11 oxidation level on their biological activity,¹⁰ we explored efficient synthetic routes to irciniastatins. Herein, we provide a full account of our work including the total synthesis of (+)-irciniastatin A (1) and (-)-irciniastatin B (2). Subsequently, we describe the binding mode of irciniastatins on the ribosome, which might induce the inhibition of protein translation.

RESULTS AND DISCUSSION

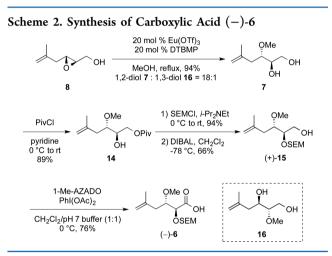
Total Synthesis of (+)-Irciniastatin A (1). Considering stereochemical complexity and structural clarity, coupled with our assumption that irciniastatins would be only distinguished by the oxidation stage at C11, we set (+)-irciniastatin A as the first goal of our synthetic venture. Our first-generation retrosynthetic analysis of (+)-irciniastatin A (1) is depicted in Scheme 1. 1 was disconnected at the amide bond, leading to the protected hemiaminal 5 and the C1–C6 acyclic side chain 6. For the installation of the hemiaminal functionality in 5, we relied on the Curtius rearrangement approach developed by Smith and co-workers.^{4b,9} The key intermediate 5 was

Scheme 1. First-Generation Retrosynthetic Analysis of (+)-Irciniastatin A (1)



conceived to be transformed from 9 via a diastereoselective reduction of the aldol moiety and a reductive translactonization. We envisioned that 9 would be united via a substrate-controlled aldol reaction of the C16-C25 ketone fragment 10 and the C8-C15 tetrahydropyran fragment 11. We envisioned synthesizing not only both fragments 6 and 11 but also their enantiomers and diastereomers for future SAR study; we decided to adopt the "Sharpless asymmetric epoxidationregioselective epoxide ring opening" sequence as a key maneuver.¹¹ 1,2-Diol 7 would be synthesized via a Lewis acid mediated regioselective ring opening from 2,3-epoxy alcohol 8, which could be easily obtained using the Sharpless asymmetric epoxidation reaction. Independently, tetrahydropyran unit of 11 would be constructed in the same manner from epoxy alcohol 13. The aldol counterpart 10 would be accessed from *E*-alkene 12 via the Sharpless asymmetric dihydroxylation, γ lactonization, and oxidation of the resultant alcohol.

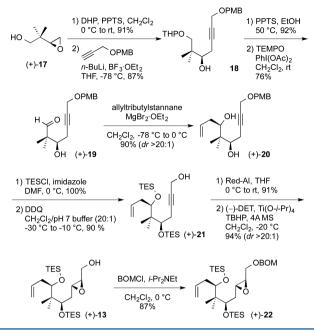
The synthesis of side chain 6 began with the regioselective ring opening of the known 2,3-epoxy alcohol (+)- 8^{12} with MeOH (Scheme 2). The Ti(O-*i*-Pr)₄-mediated nucleophilic



ring opening of 2,3-epoxy alcohols was developed by Sharpless and co-workers, which provides 3-substituted-1,2-diol derivatives.¹³ Initially, we tried this condition, but the yield and selectivity were unsatisfactory, giving the desired 1,2-diol 7 and undesired 1,3-diol **16** (54% yield, 7:**16** = 3:1) as an inseparable mixture. After extensive screening of the reaction conditions, we found that the combined use of catalytic amounts of Eu(OTf)₃ and 2,6-di-*tert*-butyl-4-methylpyridine (DTBMP) gave the desired 1,2-diol 7 in high yield and regioselectivity. In addition, we also found that this catalytic method can be applied to various epoxy alcohols and nucleophiles (alcohols, thiols, and amines).¹⁴ The 1,2-diol 7 was converted to primary alcohol (+)-**15** via a protection–deprotection sequence. Oxidation of (+)-**15** using 1-Me-AZADO/PhI(OAc)₂¹⁵ provided carboxylic acid (–)-**6**.

Access to the 2,6-trans-tetrahydropyran fragment 11 began with the known epoxy alcohol (+)-17,¹⁶ which was prepared from (-)-pantolactone (Scheme 3). Protection of the primary alcohol (+)-17 as the THP ether, followed by regioselective ring opening of epoxide with lithium acetylide and BF₃·OEt₂,¹⁷ furnished the secondary alcohol 18. Removal of the THP group and the subsequent TEMPO-catalyzed oxidation gave aldehyde (+)-19 selectively. A diastereoselective allylation of (+)-19 using allyltributylstannane in the presence of MgBr₂¹⁸ produced diol (+)-20 with high selectivity. Protecting group manipu-

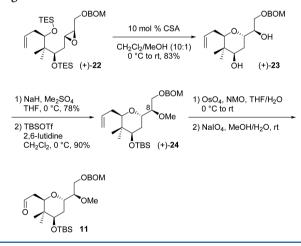
Scheme 3. Synthesis of Tetrahydropyran Precursor (+)-22



lation, followed by (E)-selective reduction of the alkyne, furnished the corresponding allyl alcohol, which was then converted to the epoxide (+)-22 through the Sharpless asymmetric epoxidation and protection of the primary alcohol. We then focused on the epoxide ring opening-tetrahy-

dropyran ring formation using (+)-22 (Scheme 4). Fortunately,

Scheme 4. Concise Synthesis of 2,6-*trans*-Tetrahydropyran Fragment 11



we found that the catalytic amount of camphorsulfonic acid (CSA) simultaneously promoted the removal of the TES group and oxy-cyclization to furnish the 2,6-*trans*-tetrahydropyran (+)-23.¹⁹ For regioselective methylation of the C8 hydroxyl group, four types of conditions were screened: (1) KOH/MeI (64% yield), (2) *t*BuOK/MeI (39% yield), (3) DTBMP/ MeOTf (complex mixtures), and (4) NaH/Me₂SO₄ (78%), indicating that bulkiness of the methylating reagent was important. Protection of the C11 hydroxyl group and the oxidative cleavage of the terminal alkene gave the desired aldehyde 11.

Synthesis of the requisite left-wing fragment (+)-10 began with the installation of a methyl group onto 26 to give 27 via an

electrophilic methoxymethylation of the benzene nuclei and the following hydrogenolytic cleavage of the methoxy group (Scheme 5). The remaining phenolic group was triflated to give 28, the Suzuki–Miyaura coupling of which with alkenyl boronate 29 in the presence of Pd(PPh₃)₄ and K₃PO₄ in dioxane at 100 °C gave 12 in 83% yield. Upon treatment with Sharpless AD-mix- α , the *E*-alkene 12 furnished γ -lactone (+)-30 in 88% (>99% ee after recrystallization) via the enantioselective dihydroxylation and concomitant lactonization. 1-Me-AZADO-catalyzed oxidation¹⁵ of (+)-30 using PhI-(OAc)₂ gave ketone (+)-10 without losing enantiomeric integrity.

Having secured both the coupling partners, we next examined the aldol reaction to construct the C15–C16 bond. Disappointingly, our attempt was hampered by the highly acidic nature of the C18-methine proton, which caused a facile scrambling of enolates generated from (+)-10, impairing the enantiomeric purity. Worse still, a model study using nonanal with lithium enolate, generated *in situ* from the ketone (+)-10 and LDA in THF at -78 °C, gave exclusively C18-aldol 31, indicating the pronounced reactivity of the C18-enolate in the aldol reaction compared with the C16-enolate. These observations led us to alter our synthetic plan shown in Scheme 6.

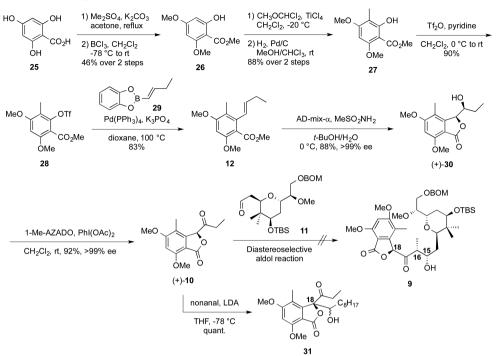
Thus, the projected key intermediate **5** was disconnected in a retro-aldol fashion into the C17–C25 aldehyde fragment **32** and the C8–C16 tetrahydropyran fragment **33** in light of successful precedents by De Brabander^{3,4g} and Smith.^{4b,9} The requisite ketone **33** would be prepared from the aldehyde **11** via nucleophilic introduction of an ethyl group.

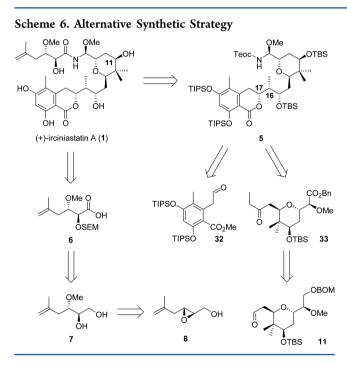
To install the ethyl ketone moiety, **11** was treated with ethylmagnesium bromide, followed by 1-Me-AZADO oxidation,¹⁵ to give ketone (+)-**34** in good yield (Scheme 7). To complete the synthesis of the tetrahydropyran fragment, one-pot oxidation conditions developed in our laboratory²⁰ were utilized for the construction of the carboxylic acid moiety. Thus, deprotection of the BOM ether gave the corresponding primary alcohol, which was oxidized using 1-Me-AZA-DO⁺BF₄⁻/NaClO₂, followed by treatment with benzyl bromide, to provide the desired central fragment (+)-**33**.

With the fragment (+)-33 in hand, we examined the coupling reaction of (+)-33 with the aldehyde 32.³ Treatment of (+)-33 with dichlorophenylborane²¹ provided the Z-enolate, and the addition of 32 provided the aldol product (+)-35 with high diastereoselectivity (Scheme 8).²² Subsequent diastereoselective reduction of the C15 ketone (+)-35 with NaBH₄ in the presence of Et₃B, followed by lactonization, provided dihydroisocoumarin (+)-36. The benzyl ester moiety in (+)-36 was then converted to a carboxylic acid via hydrogenation, which was subjected to a Curtius sequence using 2-(trimethylsilyl)ethanol as a nucleophile to give Teoc-protected hemiaminal (+)-5 in high yield.

For the synthesis of 1, the stage was set to examine the coupling of (+)-5 with the acylic chain (-)-6. Initially, we tried the coupling using the acyl chloride derivative from (-)-6, but we only observed decomposition of acid chloride. We assumed that these results were due to instability of acid chloride compounds in this condition. After intensive effort, we found that the choice of the protecting group of alcohol in (-)-6 was important,²³ and Smith's protocol^{4b,9} was the only successful method of this coupling reaction. Finally, the deprotection of all of the protecting groups using TASF provided (+)-irciniastatin A [(+)-1] in 53% yield. We used this synthetic route for the

Scheme 5. Synthesis of Left Fragment (+)-10 and Attempts at Aldol Reaction

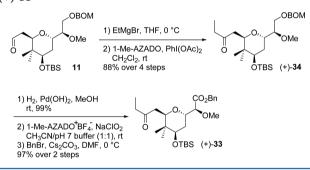




synthesis of unnatural (-)-irciniastatin A [(-)-1] and (+)-alkymberin [(+)-39] for our biological studies.^{4d}

Total Synthesis of (–)-Irciniastatin B (2). Toward the total synthesis of (–)-irciniastatin B (2), we initially ambitioned the direct conversion strategy from (+)-irciniastatin A [(+)-1] (Scheme 9). Thus, we anticipated that the C11 hydroxyl group would be the most sterically hindered among the hydroxyl groups of irciniastatin A (1), thereby resisting protection by a hindered silyl group. Indeed, the treatment of (+)-1 with 7 equiv of TBSOTf gave tetrakis-TBS ether together with pentakis-TBS ether. After chromatographic separation, the tetrakis-TBS ether was oxidized by AZADO¹⁵ to furnish the ketone. However, extensive structural analysis revealed that the

Scheme 7. Concise Synthesis of Tetrahydropyran Fragment (+)-33

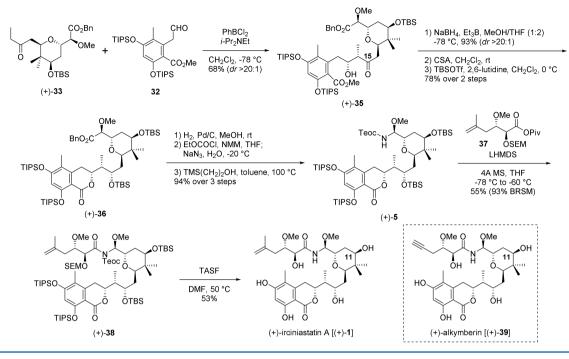


product was not **41** but (+)-**40** having the unexpected C15carbonyl moiety instead of C11.²⁴ This result could be due to the decrease in the nucleophilicity of the C15-hydroxy group via the intramolecular hydrogen bonding effect. Because the direct conversion from irciniastatin A to B was difficult, we decided to develop an alternative strategy toward the synthesis of **2**.

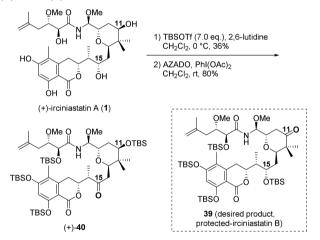
For the synthesis of (-)-irciniastatin B (2), we considered that the oxidation of C11 should be conducted at the late stage of the synthesis of **2**. In fact, Smith and co-workers reported that the 2,6-trans-tetrahydropyran-4-one core was very unstable under basic conditions.⁶ The synthesis of new tetrahydropyran fragment (+)-44 for (-)-irciniastatin B (2) commenced from the functional group manipulation of diol (+)-23 (Scheme 10). Upon the selective methylation of the C8 hydroxy group, and the deprotection of the benzyloxymethyl group using LiBF₄, ²⁵ (+)-23 was converted to diol (+)-42, the remaining two hydroxyl groups of which were protected by benzyl ether to give (-)-43 in 80% yield. Next, the ketone (+)-44 was synthesized via a four-step sequence, as in the synthesis of (+)-33.

With (+)-44 in hand, we examined the aldol coupling with the aldehyde 32 (Scheme 11). Also in this case, dichlor-

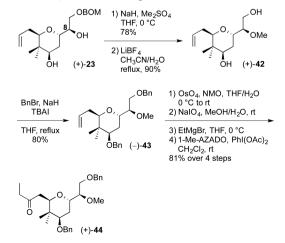




Scheme 9. Attempted Direct Conversion of Irciniastatin A (1) to B (2)



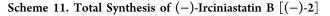




ophenylborane²¹ was the best Lewis acid for the coupling, which gave (+)-45 with good diastereoselectivity. Reduction of (+)-45 with NaBH₄ in the presence of Et₂BOMe provided the 1,3-syn diol product, which was converted to lactone (+)-46 in 78% over 2 steps. We then focused on the selective oxidation of the primary alcohol to the corresponding carboxylic acid. After screening, we found that the selective one-pot oxidation proceeded smoothly using the salt of DMN-AZADO²⁶ (1,5dimethyl-9-azanoradamantane N-oxyl), which gave (+)-48 in 81% yield. Note that the same oxidative conversion using TEMPO⁺BF₄⁻ catalyst resulted in a modest yield of (+)-48 (12) h, 56% yield). Subsequent Curtius rearrangement, followed by the protection of the C11 alcohol as a TES ether, furnished Teoc-protected hemiaminal (+)-50. The final fragment union of (+)-50 with the acyclic side chain 37 was achieved using the same conditions as those for the synthesis of (+)-38. The selective deprotection of the TES ether of (+)-51 using 1 M HCl, followed by AZADO oxidation,¹⁵ gave protected irciniastatin B (+)-52. Finally, global deprotection using TASF provided (-)-irciniastatin B (2). However, there were some slight differences in ¹H and ¹³C NMR spectra and the optical rotation value between the (-)-2 isolated by Pettit and co-workers¹ and our synthetic (-)-2.

Nevertheless, all of the spectral data for our synthetic (-)-2 were in full agreement with the data reported by Smith and coworkers.⁹ Thus, we concluded that the structure and absolute configuration of our synthetic (-)-2 are correct. In addition, we confirmed that our synthetic (-)-2 possessed the desired 2,6*trans*-tetrahydropyran-4-one core by 2D-NOE correlations.²⁷

Biological Studies of Irciniastatins A and B. Synthetic irciniastatins A and B were evaluated for their cytotoxicity against mammalian cell lines (Table 1). As we previously reported,²⁸ (+)-irciniastatin A [(+)-1] showed potent cytotoxicity, but (-)-irciniastatin B [(-)-2] showed more potent cytotoxicity. We previously reported that (+)-irciniastatin A [(+)-1] inhibited protein translation and cell cycle progression in the G1 phase in Jurkat cells. As expected, both irciniastatins



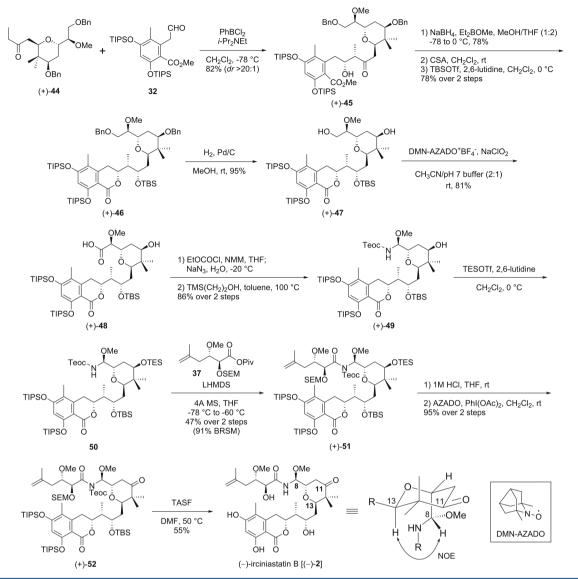


Table 1. IC₅₀ Value for Cytotoxicity against Mammalian Cells^a

	IC ₅₀ (nM)				
	HeLa	3Y1	MCAS	SKOV3	
irciniastatin A	0.49 ± 0.10	1.24 ± 0.40	1.36 ± 0.37	0.34 ± 0.03	
irciniastatin B	0.25 ± 0.02	0.41 ± 0.04	0.61 ± 0.08	0.16 ± 0.00	

^aExponentially growing cells were treated with various concentrations of irciniastatins A and B for 48 h (HeLa and 3Y1) or 72 h (MCAS and SKOV3). Cell viability was determined by WST-8. Values are expressed as mean \pm deviation of two independent experiments (MCAS and SKOV3), or mean \pm SD of three independent experiments (HeLa and 3Y1).

A and B also potently inhibited protein translation in HeLa and 3Y1 cells and cell cycle progression in G1 phase in 3Y1 cells (Tables 2 and 3). These results strongly suggest that (-)-irciniastatin B [(-)-2] as well as (+)-irciniastain A [(+)-1] is a potent protein translation inhibitor.

Chemical footprinting has been used to map the binding site of translation inhibitors, and mycalamide B (4), another pederin-type translation inhibitor, has given rise to a footprint at the E-site of the 60S ribosomal subunit.²⁹ Therefore, we next investigated whether (+)-irciniastatin A [(+)-1] and (-)-irciniastatin B [(-)-2] also bind in the same site to the mycalamide B-binding site.

Purified yeast ribosomes were incubated with each compound, followed by treatment with dimethyl sulfate (DMS). Footprints were obtained with extracted ribosome RNA by primer extension using avian myeloblastosis virus (AMV) reverse transcriptase. As shown in Figure 2, a specific methylation of C2765 on the 25S rRNA (corresponding to C3993 on 28S rRNA in mammalian cells) of the large subunit by DMS treatment was seen in DMSO samples, and this methylation was inhibited by pretreatment with 100 nM (+)-irciniastatin A [(+)-1], (-)-irciniastatin B [(-)-2], and cycloheximide (CHX). These results strongly suggested that (+)-irciniastatin A [(+)-1] and (-)-irciniastatin B [(-)-2] bind

Table 2. IC_{50} Value for Protein Synthesis of 3Y1 and HeLa Cells^{*a*}

	IC ₅₀	(nM)
	HeLa	3Y1
irciniastatin A	3.84 ± 0.13	3.05 ± 0.33
irciniastatin B	0.98 ± 0.06	0.71 ± 0.04

^{*a*}Exponentially growing 3Y1 and HeLa cells were treated with various concentrations of irciniastatins A and B for 2 h, followed by [methyl-³H] methionine treatment for 2 h. Radioactivity in acid-insoluble fractions was determined. Values are expressed as mean \pm deviation of two independent experiments.

Table 3. Distribution of DNA Content in 3Y1^a

		DNA content (%)		
	concentration (nM)	2C	2-4C	4C
control	0.0	63.7	25.6	10.7
irciniastatin A	1.0	78.5	15.5	6.0
irciniastatin B	0.3	77.0	16.9	6.1

^{*a*}Exponentially growing 3Y1 cells were treated with irciniastatins A and B at indicated concentrations for 18 h, and the distribution of DNA content and relative cell number were determined.

in the same site to the mycalamide B-binding site, the E-site of ribosome.

Taken together, the results strongly suggest that (+)-irciniastatin A [(+)-1] and (-)-irciniastatin B [(-)-2] bind in the E-site of the 60S ribosomal subunit and inhibit protein translation.

CONCLUSION

We have accomplished the divergent total synthesis of (+)-irciniastatin A [(+)-1] and (-)-irciniastatin B [(-)-2]using "Sharpless asymmetric epoxidation-regioselective epoxide ring opening" chemistry as a key maneuver. In the synthesis of the side chain, we have demonstrated an effective protocol for regioselective epoxide ring opening reaction using Eu- $(OTf)_3/DTBMP$, which realized the shortest (eight steps from commercially available propargyl alcohol) synthesis of this fragment reported to date. The convergent synthetic routes were developed via a late-stage assembly of C1-C6, C8-C16, and C17-C25 fragments. In the total synthesis of (-)-irciniastatin B, we achieved the C11-selective control of the oxidation stage via expedient functional group transformations and AZADO-catalyzed alcohol oxidation. Eight of nine stereocenters (for irciniastatin A)/seven of eight stereocenters (for irciniastatin B), except for one derived from (-)-pantolactone, were constructed employing Sharpless asymmetric epoxidation or substrate control transformations. Although our synthetic routes rely on frequent use of protecting groups (for irciniastatin A, 11 steps out of a total of 31 steps were allocated for protection/deprotection reaction; for irciniastatin B, 12 steps out of a total of 33 steps), the robust sequence allowed giving a sufficient amount of irciniastatins for biological evaluation.

The biological studies using synthetic compounds showed a high level of cytotoxic activity against mammalian cells. Furthermore, chemical footprinting experiments revealed that the binding site of irciniastatins is the E-site of the ribosome.

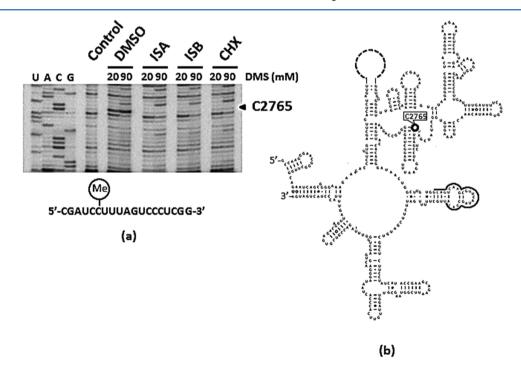


Figure 2. Inhibition of protein synthesis by binding to the E-site of the 60S subunit: The chemical footprinting of irciniastatins A and B. Ribosomes were incubated with 100 nM solutions of the compounds as indicated and methylated with DMS. Extracted rRNAs were subjected to reverse transcription. (a) rRNA not treated with DMS was the control. The $[^{32}P]$ -labeled DNA was resolved on a denaturing polyacrylamide gel. (b) Secondary structure of the 25S LSU domain V (nucleotides 2365–2423 and 2607–2994). The primer hybridized with extracted rRNA is underlined. The binding site (C2765) in domain V of the 25S rRNA is indicated.

EXPERIMENTAL SECTION

General Procedure. All reactions were carried out under an argon atmosphere with dehydrated solvents under anhydrous conditions, unless otherwise noted. Dehydrated THF and CH2Cl2 were purchased and other solvents were dehydrated and distilled according to standard protocols. Reagents were obtained from commercial suppliers and used without further purification, unless otherwise noted. Reactions were monitored by thin-layer chromatography (TLC) carried out on silica gel plates (60F₂₅₄). Column chromatography was performed on Silica gel 60N (spherical, neutral, 63-210 μ m), and flash column chromatography was performed on Silica gel 60N (spherical, neutral, $40-50 \mu$ m). Optical rotations were measured at rt. using the sodium D line. Infrared spectra were obtained at 4.0 cm⁻¹ resolution and are reported in wavenumbers. Proton nuclear magnetic resonance (¹H NMR) spectra were recorded using 400 or 600 MHz spectrometers. The chemical shifts (δ) are given from TMS (0.00 ppm) in CDCl₃ and from the residual nondeuterated solvent peak in methanol-d4 (methanol-d4:3.30 ppm) as internal standards. Coupling constants (J) are reported in hertz. The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q =quartet, m = multiplet, sept = septet, br = broad. Carbon-13 nuclear magnetic resonance (¹³C NMR) spectra were recorded using a 100 or 150 MHz spectrometer. The chemical shifts are given from CDCl₃ (77.0 ppm) and methanol-d4 (49.0 ppm) as internal standards. Lowresolution mass spectra (MS) and high-resolution mass spectra (HRMS) were recorded using electron impact (EI) with a magnetic sector or time-of-flight mass analyzer, or by fast atom bombardment (FAB) with a magnetic sector or time-of-flight mass analyzer, or by electrospray ionization (ESI) with an ion-trap mass analyzer. HPLC was performed using a UV/vistbl1 detector at 254 nm.

(2*R*,3*S*)-3-Methoxy-5-methylhex-5-ene-1,2-diol 7. To a solution of epoxy alcohol 8^{12} (708 mg, 5.53 mmol) in MeOH (28 mL) was added 2,6-di-*tert*-butyl-4-methylpyridine (227 mg, 1.11 mmol), followed by Eu(OTf)₃ (780 mg, 1.11 mmol), and the reaction was stirred for 19 h at 70 °C. After cooling of the reaction, sat. NaHCO₃ (15 mL) was added and the layers were separated. The aqueous layer was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 2/1) gave diol 7 (835 mg, 5.22 mmol, 94%) as a pair of regioisomers (1,2-diol 7:1,3-diol 16 = 18:1 determined by ¹H NMR spectrum).

7: IR (neat): 3390, 2936, 1648, 1446, 1214, 1100 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 4.84 (brs, 1H), 4.80 (brs, 1H), 3.84–3.77 (m, 1H), 3.73–3.67 (m, 2H), 3.54 (dt, *J* = 6.6, 4.1 Hz, 1H), 3.47 (s, 3 × 1/19H), 3.43 (s, 3 × 18/19H), 2.70 (d, *J* = 6.3 Hz, 1H), 2.44–2.41 (m, 1H), 2.38 (dd, *J* = 14.1, 6.8 Hz, 1H), 2.18 (dd, *J* = 14.3, 6.5 Hz, 1H), 1.79 (s, 3H); MS (EI) calculated for C₇H₁₃O₂ [M – CH₃O]⁺: 129.0916, found 129.0894.

(2*R*,3*S*)-2-Hydroxy-3-methoxy-5-methylhex-5-en-1-yl Pivalate 14. To a solution of diol 7 (205 mg, 1.28 mmol) in pyridine (2.6 mL) was added PivCl (0.173 mL, 1.41 mmol) at 0 °C. The reaction was allowed to warm to rt, and after 2 h, the reaction was cooled to 0 °C and Et₂O (3 mL) was added. Sat. NaHCO₃ (3 mL) was then added, and the mixture was extracted with Et₂O. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/8) gave pivalate ester 14 (276 mg, 1.13 mmol, 89%) as a pair of regioisomers.

14: IR (neat): 3479, 2973, 2828, 1730, 1460, 1285, 1164, 1104 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 4.78 (s, 1H), 4.76 (s, 1H), 4.18 (dd, *J* = 11.2, 3.6 Hz, 1H), 4.11 (dd, *J* = 11.2, 6.4 Hz, 1H), 3.85–3.82 (m, 2H), 3.40–3.36 (m, 1H), 3.35 (s, 3H), 2.27 (dd, *J* = 14.4, 7.6 Hz, 1H), 2.21 (dd, *J* = 14.8, 5.2 Hz, 1H), 1.74 (s, 3H), 1.17 (s, 9H); MS (EI) calculated for C₁₃H₂₄O₄ [M]⁺: 244.1675, found 244.1703.

(2*R*,3*S*)-3-Methoxy-5-methyl-2-((2-(trimethylsilyl)ethoxy)methoxy)hex-5-en-1-yl Pivalate 53. To a solution of pivalate ester 14 (276 mg, 1.13 mmol) in CH_2Cl_2 (3.8 mL) was added *i*- Pr_2NEt (0.494 mL, 2.83 mmol), followed by SEMCl (0.400 mL, 2.26 mmol), at 0 °C, and the reaction was allowed to warm to rt. After stirring for 4 h, the reaction was cooled to 0 °C and sat. NH_4Cl (3 mL) was added. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over $MgSO_{4}$, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/20) gave SEM-ether **53** (399 mg, 1.07 mmol, 94%) as a pair of regioisomers.

53: IR (neat): 2955, 1733, 1249, 1159, 1106, 1059, 1031 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 4.82 (brs, 1H), 4.79 (brs, 1H), 4.77 (d, *J* = 7.2 Hz, 1H), 4.74 (d, *J* = 7.2 Hz, 1H), 4.30 (dd, *J* = 11.6, 4.0 Hz, 1H), 4.12 (dd, *J* = 11.6, 6.4 Hz, 1H), 3.83 (dt, *J* = 6.4, 4.0 Hz, 1H), 3.69 (dt, *J* = 10.0, 6.4 Hz, 1H), 3.60 (dt, *J* = 10.0, 6.8 Hz, 1H), 3.50 (ddd, *J* = 7.2, 5.2, 4.0 Hz, 1H), 3.40 (s, 3H), 2.28–2.26 (m, 2H), 1.78 (s, 3H), 1.20 (s, 9H), 0.95–0.90 (m, 2H), 0.00 (s, 9H); MS (FAB) calculated for C₁₉H₃₉O₅Si [M + H]⁺: 375.2548, found 375.2567.

(2*R*,3*S*)-3-Methoxy-5-methyl-2-((2-(trimethylsilyl)ethoxy)methoxy)hex-5-en-1-ol (+)-15. To a solution of SEM ether 53 (399 mg, 1.07 mmol) in CH₂Cl₂ (5.3 mL) was added DIBAL (1.0 M in toluene, 2.35 mL, 2.35 mmol) at -78 °C. After 10 min, the reaction was quenched with MeOH (0.6 mL). The reaction was allowed to warm to rt, and EtOAc (6 mL) and sat. Rochelle's salt (6 mL) were added. After 1 h, the layers were separated and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/6) gave alcohol (+)-15 (204 mg, 0.704 mmol, 66%) as a colorless oil.

(+)-15: $[\alpha]_{23}^{23}$ +34.3 (c 1.98, CHCl₃); IR (neat): 3454, 2952, 1649, 1445, 1376, 1250, 1107 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 4.80 (s, 1H), 4.80 (d, *J* = 7.2 Hz, 1H), 4.76 (s, 1H), 4.68 (d, *J* = 6.8 Hz, 1H), 3.77–3.68 (m, 3H), 3.68–3.55 (m, 2H), 3.48 (dt, *J* = 9.6, 4.4 Hz, 1H), 3.9 (s, 3H), 3.24 (dd, *J* = 7.2, 4.4 Hz, 1H), 2.30 (dd, *J* = 14.4, 7.6 Hz, 1H), 2.20 (dd, *J* = 14.8, 5.2 Hz, 1H), 1.76 (s, 3H), 0.94 (t, *J* = 8.4 Hz, 2H), 0.00 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ : 142.3, 112.9, 95.3, 82.2, 80.7, 65.7, 62.3, 58.3, 39.2, 22.7, 18.0, -1.6 ; MS (FAB) calculated for C₁₄H₃₁O₄Si [M + H]⁺: 291.1992, found 291.1979.

(25,35)-3-Methoxy-5-methyl-2-((2-(trimethylsilyl)ethoxy)methoxy)hex-5-enoic Acid (–)-6. To a solution of alcohol (+)-15 (90.1 mg, 0.311 mmol) in CH₂Cl₂/pH 7 buffer (1.0 mL/1.0 mL) was added 1-Me-AZADO (10.3 mg, 0.0621 mmol) and PhI(OAc)₂ (300 mg, 0.932 mmol) at 0 °C. After 2 h, the reaction was quenched with sat. Na₂S₂O₃ and allowed to warm to rt. After 1 h, the mixture was extracted with EtOAc and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/1) gave carboxylic acid (–)-6 (71.5 mg, 0.235 mmol, 76%) as a colorless oil.

(-)-6: $[\alpha]_{D}^{23}$ -24.4 (c 0.53, CHCl₃); IR (neat): 2953, 1725, 1649, 1377, 1250, 1110, 1062 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 4.84 (s, 1H), 4.81 (s, 1H), 4.79 (s, 2H), 4.40 (d, *J* = 2.8 Hz, 1H), 3.75 (ddd, *J* = 8.4, 5.6, 3.2 Hz, 1H), 3.68 (dd, *J* = 9.2, 8.0 Hz, 1H), 3.44 (s, 3H), 2.40 (dd, *J* = 14.4, 8.0 Hz, 1H), 2.28 (dd, *J* = 14.4, 4.2 Hz, 1H), 1.77 (s, 3H), 0.92 (m, 2H), 0.02 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ : 174.6, 141.6, 113.3, 94.8, 80.7, 75.9, 66.1, 58.1, 38.5, 22.7, 18.0, - 1.4; MS (FAB) calculated for C₁₄H₂₈O₃SiNa [M + Na]⁺: 327.1604, found 327.1585.

2-(2-Methyl-2-((5)-oxiran-2-yl)propoxy)tetrahydro-2H-pyran 54. To a solution of the epoxide (+)-17 (16.6 g, 143 mmol) in CH_2Cl_2 (143 mL) was added DHP (16.9 mL, 186 mmol), followed by PPTS (7.17 g, 28.5 mmol), at 0 °C. After stirring for 10 h, DHP (3.90 mL, 42.8 mmol) was added, and the mixture was stirred for 1.5 h. The reaction was quenched with Et_3N and water. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over $MgSO_4$ and concentrated. Purification by column chromatography (EtOAc/Hexane 1/10) gave THP ether **54** (25.9 g, 130 mmol, 91%) as a pair of diastereomers.

54: IR (neat): 2942, 1477, 1351, 1201, 1122, 1065, 1035 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 4.59 (m, 1H), 3.87–3.81 (m, 1H), 3.61 (d, *J* = 9.2 Hz, 0.5H), 3.56 (dd, *J* = 9.2 Hz, 0.5H), 3.54–3.49 (m, 1H), 3.18 (d, *J* = 9.2 Hz, 0.5H), 3.11 (d, *J* = 9.2 Hz, 0.5H), 2.95–2.91 (m, 1H), 2.68–2.64 (m, 2H), 1.87–1.78 (m, 1H), 1.76–1.64 (m, 1H), 1.64–1.50 (m, 4H), 0.93 (s, 1.5H), 0.93 (s, 1.5H), 0.90 (s, 1.5H), 0.90 (s, 1.5H); ¹³C NMR (100 MHz, CDCl₃) δ : 98.6, 98.2, 73.9, 73.6, 61.4, 61.2, 56.9, 56.7, 43.5, 34.4, 34.4, 30.2, 30.2, 25.2, 20.2, 20.1, 20.0, 19.9,

19.0, 18.9 ; MS (EI) calculated for $C_{11}H_{19}O_3 \; [M-H]^+\!\!\!:$ 199.1334, found 199.1335.

(3*R*)-7-((4-Methoxybenzyl)oxy)-2,2-dimethyl-1-((tetrahydro-2*H*-pyran-2-yl)oxy)hept-5-yn-3-ol 18. To a solution of *p*methoxybenzyl propargyl ether (792 mg, 4.50 mmol) in THF (6.5 mL) was added *n*-BuLi (1.56 M in hexane, 2.88 mL, 4.50 mmol) at -78 °C. The reaction was allowed to stir at 0 °C for 1.5 h. The solution was cooled to -78 °C, and BF₃·OEt₂ (0.560 mL, 4.50 mmol) was added dropwise. After 30 min, epoxide 54 (300 mg, 1.50 mmol) was dissolved in THF (1 mL) and added dropwise to the reaction mixture. After 1 h, the reaction was quenched with Et₃N (1.0 mL) and sat. NaHCO₃ (5 mL) and stirred at rt for 1 h. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/4) gave alkyne 18 (491 mg, 1.31 mmol, 87%) as a pair of diastereomers.

18: IR (neat): 3467, 2942, 2871, 1612, 1514, 1249, 1074, 1034 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.28 (d, *J* = 8.8 Hz, 2H), 6.87 (d, *J* = 8.8 Hz, 2H), 4.59 (t, *J* = 3.4 Hz, 0.5H), 4.55–4.53 (m, 2.5H), 4.16–4.14 (m, 2H), 3.86–3.79 (m, 4H), 3.72–3.68 (m, 1.5H), 3.60 (d, *J* = 9.5 Hz, 0.5H), 3.54–3.51 (m, 1H), 3.35 (brs, 0.5H), 3.28 (d, *J* = 9.8 Hz, 0.5H), 3.19 (d, *J* = 9.5 Hz, 0.5H), 3.13 (brs, 0.5H), 2.56–2.46 (m, 1H), 2.42–2.34 (m, 1H), 1.79–1.68 (m, 2H), 1.62–1.51 (m, 4H), 0.96 (s, 1.5H), 0.95 (s, 1.5H), 0.94 (s, 1.5H), 0.93 (s, 1.5H); ¹³C NMR (100 MHz, CDCl₃) δ : 159.3, 129.7, 129.7, 128.6, 113.9, 113.8, 99.4, 99.0, 85.1, 84.9, 76.2, 75.9, 75.8, 75.7, 71.1, 71.1, 65.0, 62.7, 62.0, 57.4, 57.4, 55.3, 55.2, 38.3, 38.1, 30.6, 30.4, 25.3, 25.3, 22.9, 22.6, 22.4, 22.4, 19.7, 19.6, 19.2, 19.2; MS (EI) calculated for C₁₇H₂₃O₄ [M – C₅H₄O]⁺: 291.1596, found 291.1582.

(*R*)-7-((4-Methoxybenzyl)oxy)-2,2-dimethylhept-5-yne-1,3diol (+)-55. To a solution of alkyne 18 (3.18 g, 8.45 mmol) in EtOH (42 mL) was added PPTS (425 mg, 1.69 mmol), and the reaction was stirred for 19 h at 70 °C. After cooling of the reaction, sat. NaHCO₃ (20 mL) was added and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/2) gave diol (+)-55 (2.27 g, 7.78 mmol, 92%) as a colorless oil.

(+)-55: $[\alpha]_{2}^{28}$ +24.4 (c 1.18, CHCl₃); IR (neat): 3411, 2958, 2225, 1612, 1514, 1250, 1066 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.27 (d, *J* = 8.5 Hz, 2H), 6.88 (d, *J* = 8.5 Hz, 2H), 4.51 (s, 2H), 4.14 (t, *J* = 2.2 Hz, 1H), 3.80 (s, 3H), 3.69 (dd, *J* = 9.5, 3.2 Hz, 1H), 3.52 (d, *J* = 11.0 Hz, 1H), 3.44 (d, *J* = 11.0 Hz, 1H), 3.05 (brs, 1H), 2.92 (brs, 1H), 2.49 (ddt, *J* = 16.7, 3.2, 2.2 Hz, 1H), 2.38 (ddt, *J* = 16.7, 9.5, 2.2 Hz, 1H), 0.90 (s, 3H), 0.89 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 159.3, 129.6, 129.4, 113.8, 84.2, 78.3, 76.9, 71.6, 71.3, 57.3, 55.2, 38.3, 23.0, 22.2, 18.5 ; MS (EI) calculated for C₁₇H₂₄O₄ [M]⁺: 292.1675, found 292.1659.

(*R*)-3-Hydroxy-7-((4-methoxybenzyl)oxy)-2,2-dimethylhept-5-ynal (+)-19. To a solution of diol (+)-55 (20.0 g, 68.5 mmol) in CH_2Cl_2 (69 mL) was added PhI(OAc)_2 (28.7 g, 89.0 mmol), followed by TEMPO (1.61 g, 10.3 mmol). After stirring for 7 h at rt, PhI(OAc)_2 (4.41 g, 13.7 mmol) and TEMPO (530 mg, 3.42 mmol) were added, and the reaction was stirred for 1 h. The reaction was then diluted with Et_2O (70 mL) and quenched with sat. $Na_2S_2O_3$ (70 mL) at 0 °C. After stirring for 1 h, the mixture was extracted with Et_2O and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/3) gave aldehyde (+)-19 (15.1 g, 51.9 mmol, 76%) as a colorless oil.

(+)-19: $[\alpha]_{25}^{25}$ +24.3 (c 1.38, CHCl₃); IR (neat): 3438, 2937, 2235, 1723, 1612, 1250, 1070 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 9.57 (s, 1H), 7.27 (d, *J* = 8.4 Hz, 2H), 6.89–6.86 (m, 2H), 4.51 (s, 2H), 4.13 (t, *J* = 2.2 Hz, 1H), 3.92 (dd, *J* = 9.3, 3.7 Hz, 1H), 3.81 (s, 3H), 2.50 (ddt, *J* = 16.7, 3.7, 2.2 Hz, 1H), 2.39 (ddt, *J* = 16.7, 9.3, 2.2 Hz, 1H), 1.11 (s, 3H), 1.09 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 205.3, 159.3, 129.6, 129.4, 113.8, 83.1, 79.0, 73.4, 71.3, 57.2, 55.2, 49.7, 22.9, 18.8, 16.7 ; MS (EI) calculated for C₁₇H₂₂O₄ [M]⁺: 290.1518, found 290.1504.

(4*R*,6*R*)-10-((4-Methoxybenzyl)oxy)-5,5-dimethyldec-1-en-8yne-4,6-diol (+)-20. To a solution of MgBr·OEt₂ (405 mg, 1.57 mmol) in CH₂Cl₂ (2.0 mL) was added aldehyde (+)-19 (207 mg, 0.713 mmol) in CH₂Cl₂ (1.5 mL) at -78 °C. After stirring for 30 min, allyltributylstannane (0.490 mL, 1.57 mmol) was added, and the reaction was allowed to stir at 0 °C. After 12 h, the reaction was quenched with sat. NaHCO₃ (3 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/3) gave 4*R*-alcohol (+)-20 (214 mg, 0.644 mmol, 90%, dr > 20:1) as a colorless oil.

(+)-20: $[\alpha]_{24}^{24}$ +30.0 (c 1.74, CHCl₃); IR (neat): 3399, 2965, 2225, 1612, 1513, 1250, 1062 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ : 7.27 (d, *J* = 8.5 Hz, 2H), 6.87 (d, *J* = 8.5 Hz, 2H), 5.92–5.82 (m, 1H), 5.18–5.14 (m, 2H), 4.51 (s, 2H), 4.14 (t, *J* = 2.2 Hz, 2H), 3.81 (s, 0.06H), 3.80 (s, 2.94H), 3.76 (dt, *J* = 9.2, 3.9 Hz, 1H), 3.66 (ddd, *J* = 10.5, 3.9, 2.2 Hz, 1H), 2.49 (ddt, *J* = 16.6, 3.9, 2.2 Hz, 1H), 2.43 (ddt, *J* = 16.6, 9.0, 2.2 Hz, 1H), 2.33 (m, 1H), 2.15 (m, 1H), 0.94 (s, 3H), 0.94 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 159.1, 135.9, 129.5, 129.4, 117.5, 113.6, 84.4, 77.9, 76.7, 76.6, 71.2, 57.3, 55.2, 40.1, 36.4, 22.9, 20.8, 20.5 ; MS (FAB) calculated for C₂₀H₂₇O₄ [M – H]⁺: 331.1909, found 331.1889.

(4*R*,6*R*)-4,6-Bistriethylsilyloxy-5,5-dimethyl-10-(4-methoxybenzyloxy)-1-decen-8-yne (+)-56. To a solution of diol (+)-20 (17.2 g, 51.8 mmol) in DMF (52 mL) was added imidazole (17.6 g, 259.1 mmol), followed by TESCl (21.7 mL, 123 mmol), at 0 °C. After stirring for 1.5 h, the reaction was quenched with sat. NaHCO₃ (25 mL). The mixture was extracted with Et₂O, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/30) gave TES-ether (+)-56 (28.9 g, 51.6 mmol, 100%) as a colorless oil.

(+)-56: $[\alpha]_{25}^{25}$ +17.4 (c 1.83, CHCl₃); IR (neat): 2954, 1613, 1513, 1463, 1249, 1078, 1007 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) & 7.28 (d, *J* = 8.8 Hz, 2H), 6.90–6.86 (m, 2H), 5.85 (ddt, *J* = 18.0, 9.5, 7.2 Hz, 1H), 5.07–5.00 (m, 2H), 4.51 (s, 2H), 4.13 (t, *J* = 2.2 Hz, 2H), 3.81 (s, 3H), 3.76 (dd, *J* = 7.1, 4.8 Hz, 1H), 3.64 (dd, *J* = 7.6, 3.4 Hz, 1H), 2.50 (ddt, *J* = 17.3, 4.2, 2.0 Hz, 1H), 2.36–2.25 (m, 2H), 2.14 (m, 1H), 0.97 (t, *J* = 7.8 Hz, 9H), 0.95 (t, *J* = 7.8 Hz, 9H), 0.87 (s, 6H), 0.70 (q, *J* = 7.8 Hz, 6H), 0.60 (t, *J* = 7.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) & 159.3, 137.1, 129.7, 129.5, 126.2, 113.7, 85.8, 77.4, 77.3, 76.4, 71.0, 57.4, 55.1, 44.4, 38.0, 23.7, 19.9, 19.7, 7.0, 5.6, 5.5 ; MS (EI) calculated for C₃₀H₅₁O₄Si₂ [M - C₂H₅]⁺: S31.3326, found S31.3350.

(5*R*,*TR*)-5,7-Bistriethylsilyloxy-6,6-dimethyl-9-decen-2-yn-1ol (+)-21. To a solution of bis-TES ether (+)-56 (16.9 g, 30.1 mmol) in CH₂Cl₂/pH 7 buffer (143 mL/7.2 mL) was added DDQ (10.3 g, 45.2 mmol) at -30 °C, and the reaction was allowed to stir at 0 °C. After 13 h, the reaction was quenched with sat. Na₂S₂O₃ (100 mL) and stirred for 1 h at rt. The reaction mixture was filtered through a pad of Celite, followed by washing with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/20) gave alcohol (+)-21 (11.9 g, 27.0 mmol, 90%) as a colorless oil.

(+)-21: $[\alpha]_{25}^{25}$ +21.3 (c 1.06, CHCl₃); IR (neat): 3327, 2955, 2227, 1640, 1459, 1415, 1238, 1079, 1009 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 5.85 (ddt, *J* = 17.3, 8.9, 6.8 Hz, 1H), 5.08–5.01 (m, 2H), 4.24 (dt, *J* = 6.0, 2.2 Hz, 2H), 3.74 (dt, *J* = 7.0, 4.1 Hz, 1H), 3.64 (dd, *J* = 7.5, 3.4 Hz, 1H), 2.48 (ddt, *J* = 17.1, 4.1, 2.2 Hz, 1H), 2.33 (ddd, *J* = 14.7, 3.4, 1.7 Hz, 1H), 2.26 (ddt, *J* = 17.1, 7.0, 2.2 Hz, 1H), 2.15 (m, 1H), 1.43 (t, *J* = 6.0 Hz, 1H), 0.98 (t, *J* = 8.0 Hz, 9H), 0.96 (t, *J* = 7.3 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ : 137.2, 116.2, 85.4, 79.7, 77.4, 76.4, 51.4, 44.4, 38.0, 23.7, 20.0, 19.7, 7.1, 7.0, 7.0, 5.6, 5.4; MS (EI) calculated for C₂₂H₄₃O₃Si₂ [M - C₂H₅]⁺: 411.2751, found 411.2739.

(5*R*,7*R*,*E*)-5,7-Bistriethylsilyloxy-6,6-dimethyl-2,9-decadien-1-ol (+)-57. To a solution of alcohol (+)-21 (786 mg, 1.79 mmol) in THF (7.9 mL) was added Red-Al (1.8 M in toluene, 1.98 mL, 3.57 mmol) at 0 °C, and the reaction was allowed to stir at rt. After stirring for 3 h, the reaction was quenched with H_2O (1 mL) at 0 °C and stirred for 1 h at rt. The reaction mixture was filtered through a pad of Celite, followed by washing with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/20) gave allyl alcohol (+)-57 (718 mg, 1.62 mmol, 91%) as a colorless oil.

(+)-57: $[\alpha]_{D}^{22}$ +28.4 (c 0.65, CHCl₃); IR (neat): 3330, 2955, 2877, 1459, 1415, 1238, 1076, 1006 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 5.85 (ddt, *J* = 19.0, 10.7, 7.2 Hz, 1H), 5.78–5.61 (m, 2H), 5.05–5.01 (m, 2H), 4.11 (d, *J* = 5.6 Hz, 2H), 3.62–3.58 (m, 2H), 2.28–2.20 (m, 2H), 2.14–2.07 (m, 2H), 0.98 (t, *J* = 3.6 Hz, 18H), 0.86 (s, 6H), 0.63 (q, *J* = 3.6 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ : 137.2, 131.4, 130.5, 116.2, 77.4, 77.3, 63.8, 44.5, 37.8, 36.1, 19.8, 19.8, 7.09, 5.63 ; MS (FAB) calculated for C₂₄H₅₁O₃Si₂ [M + H]⁺: 443.3377, found 443.3394.

(2R, 3R, 5R, 7R)-5, 7-Bistriethylsilyloxy-6, 6-dimethyl-2, 3epoxy-9-decen-1-ol (+)-13. To freshly activated 4 Å molecular sieves (1.0 g) in CH₂Cl₂ (2.4 mL) was added (-)-DET (39.6 mg, 0.192 mmol) in CH_2Cl_2 (0.1 mL). The solution was cooled to -20°C, and Ti(OiPr)4 (0.0380 mL, 0.128 mmol) was added, followed by TBHP (3.11 M solution in CH₂Cl₂, 0.823 mL, 2.56 mmol). The reaction was stirred for 1 h, and then a solution of allyl alcohol (+)-57 (567 mg, 1.28 mmol) in CH₂Cl₂ (0.1 mL) was added. After 2 h, the reaction was quenched with a 6:1 mixture of acetone/water (2.8 mL) and stirred for 1 h at rt. The reaction mixture was filtered through a pad of Celite, followed by washing with EtOAc. The layers were separated, and the aqueous layer was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/12) gave epoxy alcohol (+)-13 (554 mg, 1.21 mmol, 94%, dr > 20:1) as a colorless oil.

(+)-13: $[\alpha]_D^{30}$ +46.4 (c 0.61, CHCl₃); IR (neat): 3436, 2955, 1639, 1459, 1415, 1239, 1078, 1006 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 5.84 (ddt, *J* = 15.3, 8.1, 6.8 Hz, 1H), 5.05–4.99 (m, 2H), 3.94 (ddd, *J* = 12.2, 6.1, 2.4 Hz, 1H), 3.81 (dd, *J* = 9.0, 2.4 Hz, 1H), 3.63 (ddd, *J* = 12.2, 6.1, 4.4 Hz, 1H), 3.48 (dd, *J* = 7.6, 3.4 Hz, 1H), 3.11 (dt, *J* = 7.6, 2.8 Hz, 1H), 2.94 (dt, *J* = 4.0, 2.4 Hz, 1H), 2.26–2.22 (m, 1H), 2.14–2.06 (m, 1H), 1.78 (ddd, *J* = 14.4, 9.0, 3.7 Hz, 1H), 1.62 (t, *J* = 6.1 Hz, 1H), 1.38 (ddd, *J* = 14.1, 7.8, 2.4 Hz, 1H), 0.99 (t, *J* = 7.8 Hz, 9H), 0.95 (t, *J* = 8.1 Hz, 9H), 0.88 (s, 3H), 0.85 (s, 3H), 0.66 (q, *J* = 8.1 Hz, 6H), 0.66 (q, *J* = 7.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ : 136.9, 116.2, 77.6, 75.1, 61.6, 59.7, 54.0, 44.0, 37.6, 35.1, 20.0, 19.9, 7.2, 7.2, 5.7, 5.6 ; MS (FAB) calculated for C₂₄H₅₁O₄Si₂ [M + H]⁺: 459.3326, found 459.3304.

(4*R*,6*R*,8*R*,9*R*)-10-(Benzyloxymethoxy)-4,6-bistriethylsilyloxy-5,5-dimethyl-8,9-epoxy-1-decene (+)-22. To a solution of epoxy alcohol (+)-13 (2.35 g, 5.13 mmol) in CH₂Cl₂ (10 mL) was added *i*-Pr₂NEt (3.59 mL, 20.5 mmol), followed by BOMCl (1.41 mL, 10.3 mmol), at 0 °C. After stirring for 5.5 h, the reaction was quenched with sat. NaHCO₃ (10 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/40–1/20) gave BOM ether (+)-22 (2.59 g, 4.47 mmol, 87%) as a colorless oil.

(+)-22: $[\alpha]_D^{26}$ +37.8 (c 0.49, CHCl₃); IR (neat): 2954, 1457, 1077, 1006 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) & 7.35–7.28 (m, 5H), 5.83 (ddt, *J* = 17.1, 10.0, 7.1 Hz, 1H), 5.02 (d, *J* = 15.4 Hz, 1H), 5.00 (d, *J* = 9.8 Hz, 1H), 4.78 (s, 2H), 4.62 (s, 2H), 3.84–3.80 (m, 2H), 3.58 (dd, *J* = 11.5, 5.6 Hz, 1H), 3.46 (dd, *J* = 7.6, 3.4 Hz, 1H), 3.02 (dt, *J* = 8.1, 2.9 Hz, 1H), 2.95 (ddd, *J* = 5.4, 2.9 Hz, 1H), 2.23 (m, 1H), 2.09 (m, 1H), 1.79 (ddd, *J* = 12.7, 9.3, 3.4 Hz, 1H), 1.33 (ddd, *J* = 14.2, 7.8, 2.2 Hz, 1H), 0.98 (t, *J* = 8.1 Hz, 9H), 0.95 (t, *J* = 7.8 Hz, 9H), 0.88 (s, 3H), 0.84 (s, 3H), 0.66 (q, *J* = 8.1 Hz, 6H), 0.59 (q, *J* = 7.8 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 137.7, 137.0, 128.4, 127.9, 127.7, 116.2, 94.7, 77.6, 75.1, 69.4, 68.0, 58.0, 54.2, 44.0, 37.5, 35.1, 19.9, 19.8, 7.1, 7.1, 5.6, 5.5 ; MS (EI) calculated for C₃₀H₅₃O₅Si₂ [M – C₂H₅]⁺: 549.3432, found 549.3449.

(2*R*,4*R*,65)-2-Allyl-6-((1*R*)-2-(benzyloxymethoxy)-1-hydroxyethyl)-3,3-dimethyl-4-hydroxytetrahydropyran (+)-23. To a solution of BOM ether (+)-22 (476 mg, 0.823 mmol) in $CH_2Cl_2/$ MeOH (4.1 mL/0.4 mL) was added CSA (19.0 mg, 0.0823 mmol) at 0 °C. After stirring for 19 h, the reaction was quenched with sat. NaHCO₃ (1.0 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/1) gave diol (+)-23 (238 mg, 0.680 mmol, 83%) as a colorless oil.

(+)-23: $[\alpha]_D^{26}$ +41.7 (c 0.94, CHCl₃); IR (neat): 3421, 2942, 1640, 1454, 1090 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.36–7.29 (m, SH), 5.82 (ddt, *J* = 17.1, 10.3, 6.8 Hz, 1H), 5.05 (ddd, *J* = 17.1, 3.4, 1.5 Hz, 1H), 4.99 (ddd, *J* = 10.3, 2.0, 1.0 Hz, 1H), 4.81 (d, *J* = 6.6 Hz, 1H), 4.79 (d, *J* = 6.6 Hz, 1H), 4.63 (s, 2H), 3.94 (m, 1H), 3.84 (dd, *J* = 10.6, 2.9 Hz, 1H), 3.81 (dd, *J* = 5.4, 4.0 Hz, 1H), 3.63 (dd, *J* = 9.5, 4.0 Hz, 1H), 3.59 (dd, *J* = 10.6, 6.6 Hz, 1H), 3.23 (dd, *J* = 10.7, 2.7 Hz, 1H), 2.64 (brs, 1H), 2.35 (m, 1H), 2.22 (m, 1H), 2.11 (dt, *J* = 13.4, 4.0 Hz, 1H), 1.74 (ddd, *J* = 13.4, 9.5, 5.4 Hz, 1H), 0.99 (s, 3H), 0.91 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 137.6, 136.8, 128.6, 128.0, 127.9, 116.4, 95.6, 79.5, 72.3, 70.7, 70.7, 69.9, 69.1, 38.4, 33.7, 29.7, 23.9, 14.8 ; MS (EI) calculated for C₂₀H₃₀O₅ [M]⁺: 350.2093, found 350.2108.

(2*R*,4*R*,65)-2-Allyl-6-((1*R*)-2-(benzyloxymethoxy)-1-methoxyethyl)-3,3-dimethyl-4-hydroxytetrahydropyran (+)-58. To a solution of diol (+)-23 (121 mg, 0.344 mmol) in THF (3.4 mL) was added NaH (60%, 34.4 mg, 0.861 mmol) at 0 °C. After stirring for 30 min, Me₂SO₄ (0.0390 mL, 0.413 mmol) was added, and the reaction was stirred at 0 °C. After 2 h, the reaction was quenched with sat. NH₄Cl (2 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/8–1/4) gave methyl ether (+)-58 (97.4 mg, 0.267 mmol, 78%) as a colorless oil.

(+)-**58**: $[\alpha]_D^{29}$ +33.3 (c 0.80, CHCl₃); IR (neat): 3467, 2935, 1640, 1454, 1100, 1056 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) & 7.36–7.27 (m, 5H), 5.84 (ddt, *J* = 17.1, 10.1, 7.0 Hz, 1H), 5.04 (dd, *J* = 17.1, 1.5 Hz, 1H), 4.98 (d, *J* = 10.1 Hz, 1H), 4.79 (s, 2H), 4.63 (s, 2H), 3.95 (ddd, *J* = 8.9, 6.1, 3.0 Hz, 1H), 3.90 (dd, *J* = 11.0, 2.2 Hz, 1H), 3.81 (dd, *J* = 5.4, 3.9 Hz, 1H), 3.73 (dd, *J* = 9.5, 4.2 Hz, 1H), 3.64 (dd, *J* = 11.0, 5.5 Hz, 1H), 3.59 (dd, *J* = 10.6, 4.8 Hz, 1H), 3.53 (ddd, *J* = 8.9, 4.8, 2.2 Hz, 1H), 3.47 (s, 3H), 3.20 (dd, *J* = 9.9, 2.7 Hz, 1H), 2.31–2.18 (m, 2H), 2.06 (dt, *J* = 13.4, 3.0 Hz, 1H), 1.75 (dt, *J* = 13.4, 10.6, 6.1 Hz, 1H), 1.44 (brs, 1H), 0.99 (s, 3H), 0.90 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) & 137.9, 136.7, 128.4, 127.9, 127.6, 116.3, 95.1, 79.1, 78.6, 72.5, 69.9, 69.4, 66.6, 58.4, 38.6, 33.8, 29.6, 23.5, 13.8 ; MS (EI) calculated for $C_{18}H_{27}O_5$ [M - C_3H_5]⁺: 323.1859, found 323.1841.

(2*R*,4*R*,65)-2-Allyl-6-((1*R*)-2-(benzyloxymethoxy)-1-methoxyethyl)-3,3-dimethyl-4-*tert*-butyldimethylsilyloxytetrahydropyran (+)-24. To a solution of methyl ether (+)-58 (177 mg, 0.487 mmol) in CH₂Cl₂ (2.4 mL) was added 2,6-lutidine (0.170 mL, 1.46 mmol), followed by TBSOTf (0.168 mL, 0.730 mmol), at 0 °C. After stirring for 30 min, the reaction was quenched with sat. NaHCO₃ (1 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/40–1/20) gave TBS ether (+)-24 (209 mg, 0.438 mmol, 90%) as a colorless oil.

(+)-24: $[\alpha]_{29}^{29}$ +18.5 (c 1.24, CHCl₃); IR (neat): 2954, 1640, 1471, 1102 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.36–7.27 (m, 5H), 5.83 (ddt, *J* = 16.8, 10.0, 6.8 Hz, 1H), 5.02 (d, *J* = 16.8 Hz, 1H), 4.96 (dd, *J* = 10.0, 1.0 Hz, 1H), 4.79 (s, 2H), 4.63 (s, 2H), 3.92–3.86 (m, 2H), 3.63 (dd, *J* = 10.5, 2.4 Hz, 1H), 3.56–3.50 (m, 2H), 3.49 (s, 3H), 3.23 (dd, *J* = 10.5, 2.4 Hz, 1H), 2.40 (m, 1H), 2.19 (dd, *J* = 14.4, 6.8 Hz, 1H), 1.94 (dt, *J* = 13.9, 4.4 Hz, 1H), 1.70 (ddd, *J* = 13.9, 9.3, 5.4 Hz, 1H), 1.44 (brs, 1H), 0.94 (s, 3H), 0.90 (s, 9H), 0.88 (s, 3H), 0.06 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ : 137.8, 137.0, 128.3, 127.8, 127.6, 115.9, 95.1, 79.7, 79.3, 73.0, 69.3, 69.0, 67.2, 58.5, 38.8, 33.8,

30.2, 25.9, 24.5, 18.1, 15.8, -4.1, -4.9; MS (EI) calculated for $C_{27}H_{46}O_SSi$ [M]⁺: 478.3115, found 478.3118.

2-((2R,4R,6S)-6-((R)-2-((Benzyloxy)methoxy)-1-methoxyethyl)-4-((tert-butyldimethylsilyl)oxy)-3,3-dimethyltetrahydro-2H-pyran-2-yl)acetaldehyde 11. To a solution of TBS ether (+)-24 (500 mg, 1.05 mmol) in THF/H₂O (10 mL/1 mL) was added NMO (184 mg, 1.57 mmol), followed by OsO_4 (0.196 M solution in THF, 0.0530 mL, 0.0105 mmol), at 0 °C, and the reaction was allowed to stir at rt. After stirring for 10.5 h, the reaction was quenched with sat. Na₂S₂O₃ (1 mL) and stirred for 1 h at rt. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine and dried over MgSO4. Filtration and concentration afforded a crude diol. To a solution of the diol in MeOH/H₂O (18.8 mL/2.1 mL) was added NaIO₄ (335 mg, 1.57 mmol), and the mixture was stirred at rt. After 30 min, EtOAc (30 mL) and H₂O (10 mL) were added. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine and dried over MgSO4. Filtration and concentration afforded aldehyde 11 as a colorless oil.

11: IR (neat): 2954, 1727, 1524, 1234, 1071 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 9.66 (d, J = 2.4 Hz, 1H) 7.29–7.19 (m, 5H), 4.71 (d, J = 1.2 Hz, 1H), 4.55 (d, J = 1.2 Hz, 1H), 4.61 (s, 1H), 3.89–3.84 (m, 1H), 3.72 (dd, J = 10.6, 3.0 Hz, 1H), 3.59–3.53 (m, 1H), 3.45–3.39 (m, 4H), 3.21 (d, J = 9.6 Hz, 1H), 2.87–2.80 (m, 1H), 2.42 (m, J = 16.4 Hz, 1H), 1.94–1.88 (m, 1H), 1.64–1.58 (m, 1H), 0.90 (s, 3H), 0.85 (s, 9H), 0.82 (s, 3H), 0.01 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) δ : 201.7, 137.9, 128.3, 127.8, 127.5, 95.0, 79.8, 74.9, 72.8, 69.4, 68.6, 66.7, 58.4, 43.7, 38.0, 30.1, 25.9, 24.7, 18.1, –4.2, –4.9; MS (EI) calculated for C₂₂H₃₅O₆Si [M – C₄H₉]⁺: 423.2203, found 423.2227.

Methyl 4,6-Dimethoxy-2-hydroxybenzoate 26. To a solution of 2,4,6-trihydroxybenzoic acid (**25**) (10.0 g, 53.2 mmol) in acetone (177 mL) was added K_2CO_3 (44.1 g, 319 mmol), followed by Me_2SO_4 (30.2 mL, 319 mmol), and the mixture was heated to reflux. After stirring for 22 h, the reaction mixture was filtered through a pad of Celite, followed by washing with EtOAc. Then, concentration afforded a crude trimethyl ether. To a solution of the crude trimethyl ether in CH₂Cl₂ (106 mL) was added BCl₃ (1.0 M solution, 79.8 mL, 79.8 mmol) at -78 °C, and the reaction was allowed to stir at 0 °C. After stirring for 5 h, the reaction was quenched with 10% HCl aq. and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/8) gave phenol **26** (5.16 g, 24.3 mmol, 46% over 2 steps) as a white solid.

26: mp: 111–112 °C; IR (neat): 1638 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 12.03 (s, 1H), 6.11 (d, *J* = 2.5 Hz, 1H), 5.97 (d, *J* = 2.5 Hz, 1H), 3.92 (s, 1H), 3.83 (s, 3H), 3.81 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 171.6, 166.0, 165.3, 162.1, 96.6, 93.4, 91.5, 56.0, 55.4, 52.2; MS (EI) calculated for C₁₀H₁₂O₅ [M]⁺: 212.0685, found 212.0692.

Methyl 4,6-Dimethoxy-2-hydroxy-3-methylbenzoate 27. To a solution of phenol 26 (5.16 g, 24.3 mmol) in CH_2Cl_2 (81 mL) was added 1,1-dichlorodimethyl ether (4.40 mL, 48.6 mmol), followed by TiCl₄ (8.00 mL, 72.9 mmol), at -20 °C. After stirring for 1 h, the reaction was quenched with 10% HCl aq. and the mixture was extracted with EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded a crude benzaldehyde. To a solution of the crude benzaldehyde in MeOH/CHCl₃ (54 mL/27 mL) was added 10% Pd-C (1.17 g) and hydrogenated (H₂, 1 atm). After 38 h, the catalyst was filtered through a pad of Celite, followed by washing with EtOAc. The combined organic layers were concentrated. Purification by column chromatography (EtOAc/Hexane 1/8) gave methylbenzene 27 (4.83 g, 21.4 mmol, 88%) as a white solid.

27: mp: 153–154 °C; IR (neat): 2950, 1729, 1605 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 12.04 (s, 1H), 5.99 (s, 1H), 3.92 (s, 1H), 3.87 (s, 3H), 3.86 (s, 3H), 2.03 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 171.9, 162.7, 162.1, 160.5, 105.7, 96.5, 87.0, 56.1, 55.4, 52.1, 7.4 ; MS (EI) calculated for C₁₁H₁₄O₅ [M]⁺: 226.0841, found 226.0828.

Methyl 4,6-Dimethoxy-3-methyl-2-trifluoromethanesulfonyloxybenzoate 28. To a solution of methylbenzene 27 (82.1 mg, 0.363 mmol) in CH₂Cl₂ (1.8 mL) was added pyridine (70.0 μ L, 0.871 mmol), followed by Tf₂O (0.0730 mL, 0.436 mmol), at 0 °C. After stirring for 4 h, the reaction was quenched with sat. NaHCO₃ and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/3) gave triflate **28** (117 mg, 0.327 mmol, 90%) as a white solid.

28: mp: 113–114 °C; IR (neat): 1735, 1621 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.46 (s, 1H), 3.90 (s, 3H), 3.88 (s, 3H), 2.15 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 163.8, 160.5, 157.3, 145.3, 118.3 (q, *J* = 318 Hz), 113.2, 109.5, 94.8, 56.6, 56.0, 52.5, 9.6; MS (EI) calculated for C₁₂H₁₃F₃O₇S [M]⁺: 358.0334, found 358.0336.

Methyl 2-(1-Butenyl)-4,6-dimethoxy-3-methylbenzoate 12. A solution of boronic acid ester **29** (1.14 g, 6.54 mmol) in 1,4-dioxane (16 mL) was first sonicated. To triflate **28** (1.17 g, 3.27 mmol), K_3PO_4 (1.39 g, 6.54 mmol), and Pd(PPh₃)₄ (378 mg, 0.327 mmol) was added a solution of boronic acid ester in 1,4-dioxane at rt. After stirring at 100 °C for 12 h, the reaction was quenched with 1 M NaOH aq. and the reaction mixture was filtered through a pad of Celite, followed by washing with EtOAc. The mixute was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/8) gave alkene **12** (717 mg, 2.71 mmol, 83%) as a colorless oil.

12: IR (neat): 1731, 1588 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.36 (d, *J* = 15.8 Hz, 1H), 6.36 (s, 1H), 5.76 (dt, *J* = 15.8, 6.8 Hz, 1H), 3.82 (s, 3H), 3.81 (s, 3H), 3.78 (s, 3H), 2.16 (dq, *J* = 7.6, 6.8 Hz, 3H), 1.04 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 169.1, 158.8, 155.3, 137.7, 137.6, 125.6, 116.8, 115.8, 93.8, 56.1, 55.6, 51.9, 29.3, 13.6, 11.6 ; MS (EI) calculated for C₁₅H₂₀O₄ [M]⁺: 264.1362, found 264.1351.

(35)-5,7-Dimethoxy-3-[(15)-1-hydroxypropyl]-4-methylphthalide (+)-30. To a solution of phenol AD-mix- α (530 mg, 1.4 g/ mmol) in *t*-BuOH/H₂O (1.5 mL/1.5 mL) was added MeSO₂NH₂ (36.0 mg, 0.379 mmol) at 0 °C. After stirring for 30 min, alkene 12 (100 mg, 0.379 mmol) was added, and the reaction was stirred for 108 h. Then, the reaction was quenched with sat. Na₂S₂O₃ and the crude was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/4) and then recrystallization gave phthalide (+)-30 (89.0 mg, 0.335 mmol, 88%, 99% ee) as a white solid. The enantiomeric excess was determined by HPLC analysis.

(+)-30: $[\alpha]_{D}^{31}$ +77.7 (c 1.45, CHCl₃); mp: 178–179 °C; IR (neat): 3469, 1739 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.42 (s, 1H), 5.34 (s, 1H), 4.01 (t, *J* = 7.6 Hz, 1H), 3.98 (s, 3H), 3.92 (s, 3H), 2.16 (dq, *J* = 7.6, 7.3 Hz, 3H), 2.12 (s, 3H), 1.09 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 168.9, 164.0, 158.0, 148.8, 111.9, 106.0, 94.5, 80.9, 72.1, 55.9, 55.8, 27.6, 10.9, 10.3 ; MS (EI) calculated for C₁₄H₁₈O₅ [M]⁺: 266.1154, found 266.1136.

(35)-5,7-Dimethoxy-4-methyl-3-(1-oxopropyl)-phthalide (+)-10. To a solution of phthalide (+)-30 (54.6 mg, 0.205 mmol, 99% ee) in CH₂Cl₂ (0.38 mL) was added PhI(OAc)₂ (78.7 mg, 0.244 mmol), followed by 1-Me-AZADO (7.80 mg, 0.0470 mmol), at 0 °C. After stirring for 5 h at rt, the reaction was then diluted with Et₂O (1 mL) and quenched with sat. Na₂S₂O₃ (1 mL) at 0 °C. After stirring for 1 h, the crude was extracted with EtOAc and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/2) and then recrystallization gave ketone (+)-10 (49.7 mg, 0.188 mmol, 92%, 99% ee) as a white solid. The enantiomeric excess was determined by HPLC analysis.

(+)-10: $[\alpha]_D^{24}$ +241 (c 0.66, CHCl₃); mp: 130–131 °C; IR (neat): 1768, 1726 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.46 (s, 1H), 5.58 (s, 1H), 4.01 (s, 3H), 3.94 (s, 3H), 3.81 (s, 3H), 2.61 (dq, *J* = 18.8, 7.5 Hz, 1H), 2.19 (dq, *J* = 18.8, 7.1 Hz, 1H), 2.08 (s, 3H), 0.97 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 204.8, 168.2, 164.5, 158.4, 145.0, 114.0, 103.9, 95.2, 83.3, 56.1, 56.1, 29.5, 10.7, 7.1; MS (EI) calculated for C₁₄H₁₆O₅ [M]⁺: 264.0998, found 264.0984.

(3*R*)-3-(1-Hydroxynonyl)-5,7-dimethoxy-4-methyl-3-propionylphthalide 31. To a solution of LDA in THF (1.24 mL, 0.390 mmol) was added ketone (+)-10 (87.0 mg, 0.329 mmol) in THF at

-78 °C. After stirring for 30 min, nonanal (95.0 mg, 0.658 mmol) was added. After 2 h at -78 °C, the reaction was quenched with sat. NH₄Cl at 0 °C. The crude was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/2) gave aldol **31** (134 mg, 0.329 mmol, quant., mixture of diastereomer) as a white solid.

31: IR (neat): 3464, 1759, 1725, 1614, 1597 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, major diastereomer) δ : 6.48 (s, 1H), 4.81 (dt, *J* = 9.6, 2.4 Hz, 1H), 4.02 (s, 3H), 3.95 (s, 3H), 2.69 (dq, *J* = 13.2, 5.6 Hz, 1H), 2.27 (dq, *J* = 13.2, 5.6 Hz, 1H), 2.16 (s, 3H), 2.04 (d, *J* = 9.6 Hz), 1.47–1.45 (m, 1H), 1.32–1.14 (m, 12H), 0.97 (t, *J* = 7.2 Hz, 3H), 0.84 (t, *J* = 6.4 Hz, 4H); ¹³C NMR (100 MHz, CDCl₃) δ : 208.7, 166.2, 164.8, 163.4, 160.6, 158.7, 158.1, 130.6, 121.9, 114.4, 100.0, 95.3, 83.1, 71.8, 71.5, 56.6, 56.2, 56.1, 44.8, 44.0, 33.9, 33.9, 33.5, 31.8, 29.5, 29.3, 29.2, 29.2, 29.0, 29.0, 25.9, 25.4, 24.6, 22.6, 14.0, 11.1, 10.9, 10.3, 10.0 ; MS (EI) calculated for C₂₃H₃₄O₆ [M]⁺: 406.2355, found 406.2353.

(2R,4R,6S)-6-((1R)-2-(Benzyloxymethoxy)-1-methoxyethyl)-3,3-dimethyl-2-(2-oxobutyl)-4-tert-butyldimethylsilyloxytetrahvdropyran (+)-34. To a solution of EtMgBr (0.300 M solution in THF, 20.9 mL, 6.27 mmol) was added the crude aldehyde 11 dropwise at 0 °C. After stirring for 30 min, the reaction was quenched with sat. NH₄Cl (5 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded a crude ethyl alcohol. To a solution of the ethyl alcohol in CH2Cl2 (5.2 mL) was added PhI(OAc)₂ (404 mg, 1.25 mmol), followed by 1-Me-AZADO (17.4 mg, 0.105 mmol), at rt. After stirring for 16 h, the reaction was then diluted with Et₂O (10 mL) and quenched with sat. Na₂S₂O₃ (10 mL) at 0 °C. After stirring for 1 h at rt, the mixture was extracted with EtOAc and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/10) gave ketone (+)-34 (467 mg, 0.919 mmol, 88% over 4 steps) as a colorless oil.

(+)-34: $[\alpha]_{24}^{26}$ +22.0 (c 3.60, CHCl₃); IR (neat): 2931, 1716, 1472, 1254, 1111 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.37–7.26 (m, 5H), 4.78 (d, *J* = 6.4 Hz, 1H), 4.75 (d, *J* = 6.4 Hz, 1H), 4.61 (s, 1H), 3.92–3.79 (m, 3H), 3.63–3.52 (m, 3H), 3.48 (s, 3H), 2.78 (dd, *J* = 16.0, 9.8 Hz, 1H), 2.43–2.38 (m, 3H), 1.96 (dt, *J* = 13.6, 4.0 Hz, 1H), 1.68 (ddd, *J* = 14.8, 9.6, 5.6 Hz, 1H), 1.00 (t, *J* = 7.2 Hz, 3H), 0.91 (s, 9H), 0.91 (s, 3H), 0.86 (s, 3H), 0.07 (s, 3H), 0.06 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 210.4, 138.1, 128.3, 127.9, 127.5, 95.1, 79.0, 75.7, 72.8, 69.5, 69.2, 66.3, 58.3, 42.3, 38.3, 37.3, 29.9, 25.8, 24.2, 18.0, 15.5, 7.6, -4.2, -5.0 ; MS (EI) calculated for C₂₈H₄₈O₆Si [M]⁺: 508.3220, found 508.3201.

(2*R*,4*R*,6*S*)-3,3-Dimethyl-6-((1*R*)-2-hydroxy-1-methoxyethyl)-2-(2-oxobutyl)-4-*tert*-butyldimethylsilyloxytetrahydropyran (+)-59. To a solution of ketone (+)-34 (339 mg, 0.667 mmol) in MeOH (3.3 mL) was added 20% $Pd(OH)_2$ (33.9 mg, 10% w/w) and hydrogenated (H₂, 1 atm). After 40 min, the catalyst was filtered through a pad of Celite, followed by washing with EtOAc. The combined organic layers were concentrated. Purification by column chromatography (EtOAc/Hexane 1/4) gave alcohol (+)-59 (257 mg, 0.663 mmol, 99%) as a colorless oil.

(+)-**59**: $[\alpha]_{25}^{25}$ +23.9 (c 1.59, CHCl₃); IR (neat): 3467, 2955, 1715, 1471, 1362, 1254, 1079 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 3.81 (m, 2H), 3.71 (d, *J* = 12.0 Hz, 1H), 3.60 (brs, 1H), 3.56 (dd, *J* = 8.0, 4.0 Hz, 1H), 3.42 (s, 3H), 3.29 (dt, *J* = 8.0, 3.6 Hz, 1H), 2.90 (dd, *J* = 16.0, 10.0 Hz, 1H), 2.61 (brs, 1H), 2.42 (m, 3H), 1.85 (dt, *J* = 14.0, 4.8 Hz, 1H), 1.67 (ddd, *J* = 13.6, 8.0, 4.8 Hz, 1H), 1.01 (t, *J* = 7.2 Hz, 1H), 0.90 (s, 3H), 0.87 (s, 9H), 0.83 (s, 3H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 210.1, 81.0, 79.3, 72.8, 69.0, 60.8, 58.0, 42.2, 38.0, 37.0, 30.8, 25.8, 24.7, 18.0, 17.1, 7.6, -4.3, -5.0; MS (EI) calculated for C₂₀H₄₁O₅Si [M + H]⁺: 389.2723, found 389.2761.

(2*R*,4*R*,6*S*)-6-((1*S*)-1-Benzyloxycarbonyl-1-methoxymethyl)-3,3-dimethyl-2-(2-oxobutyl)-4-*tert*-butyldimethylsilyloxytetrahydropyran (+)-33. To a solution of alcohol (+)-59 (20.9 mg, 0.0538 mmol) in MeCN/pH 7 buffer (0.27 mL/0.27 mL) was added NaClO₂ (80%, 18.3 mg, 0.161 mmol), followed by 1-Me-AZADO⁺BF₄⁻ (1.36 mg, 0.00538 mmol), at rt. After stirring for 30 min, the reaction was quenched with 2-methyl-2-butene (1 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded a crude carboxylic acid. To a solution of the crude carboxylic acid in DMF (1.1 mL) was added Cs₂CO₃ (35.1 mg, 0.108 mmol), followed by BnBr (0.0255 mL, 0.215 mmol), at 0 °C. After stirring for 1 h, the reaction was diluted with Et₂O (3 mL) and quenched with sat. NH₄Cl (2 mL). The crude was extracted with Et₂O, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/20–1/8) gave benzyl ester (+)-33 (25.6 mg, 0.0520 mmol, 97% over 2 steps) as a colorless oil.

(+)-33: $[\alpha]_{2}^{D4}$ +11.5 (c 1.30, CHCl₃); IR (neat): 2955, 1747, 1717, 1461, 1255, 1100, 1005 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.39–7.29 (m, 5H), 5.23 (d, *J* = 12.2 Hz, 1H), 5.20 (d, *J* = 12.2 Hz, 1H), 4.12 (q, *J* = 6.1 Hz, 1H), 4.03 (dd, *J* = 9.5, 3.6 Hz, 1H), 3.95 (d, *J* = 6.1 Hz, 1H), 3.64 (dd, *J* = 7.6, 3.6 Hz, 1H), 3.39 (s, 3H), 2.85 (dd, *J* = 15.4, 9.3 Hz, 1H), 2.50 (dd, *J* = 15.4, 3.4 Hz, 1H), 2.42 (q, *J* = 7.3 Hz, 2H), 1.92 (ddd, *J* = 13.9, 6.4, 3.9 Hz, 1H), 1.60 (ddd, *J* = 13.2, 7.2, 4.8 Hz, 1H), 1.02 (t, *J* = 7.3 Hz, 3H), 0.94 (s, 3H), 0.90 (s, 9H), 0.83 (s, 3H), 0.04 (s, 3H), 0.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 209.6, 170.4, 135.4, 128.3, 128.1, 128.0, 82.6, 76.8, 72.7, 69.6, 66.5, 58.3, 42.2, 37.8, 36.8, 30.0, 25.7, 24.6, 17.9, 17.3, 7.6, -4.5, -5.1; MS (EI) calculated for C₂₇H₄₄O₆Si [M]⁺: 492.2907, found 492.2911

(S)-Benzyl 2-((2S,4R,6R)-Tetrahydro-5,5-dimethyl-6-((3S,4R)-4-hydroxy-3-methyl-5-(6-methyl-2-methyloxycarbonyl-3,5bistriisopropylsilyloxyphenyl)-2-oxopentyl)-4-tert-butyldimethylsilyloxy-2H-pyran-2-yl)-2-methoxyacetate (+)-35. A solution of ketone (+)-33 (69.7 mg, 0.142 mmol) in CH₂Cl₂ (2.0 mL) was cooled to -78 °C, and PhBCl₂ (44.1 μ L, 0.340 mmol) was added. After stirring for 20 min, *i*-Pr₂NEt (74.2 μ L, 0.425 mmol) was added dropwise. After stirring for 1 h at -78 °C, aldehyde 32 (91.2 mg, 0.170 mmol) in CH₂Cl₂ (0.8 mL) was added to the boron enolate dropwise. After stirring for 1 h at -78 °C, the reaction was quenched with MeOH (3 mL) and sat. NaHCO₃ (3 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/8) gave aldol (+)-35 (99.1 mg, 0.0964 mmol, 68%, *dr* > 20:1) as a white foam.

(+)-35: $[\alpha]_{D}^{23}$ +35.2 (c 2.61, CHCl₃); IR (neat): 3452, 2947, 2867, 1730, 1589, 1469, 1259, 1166 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.38-7.29 (m, 5H), 6.26 (s, 1H), 5.24 (d, J = 12.0 Hz, 1H), 5.11 (d, J= 12.0 Hz, 1H), 4.14 (q, J = 5.6 Hz, 1H), 4.09 (dd, J = 9.2, 2.8 Hz, 1H), 4.03 (m, 1H), 3.98 (d, J = 5.6 Hz, 1H), 3.83 (s, 3H), 3.65 (dd, J = 7.6, 4.8 Hz, 1H), 3.50 (m, 1H), 3.39 (s, 3H), 3.02 (dd, J = 16.4, 9.2 Hz, 1H), 2.82 (dd, J = 14.0, 3.2 Hz, 1H), 2.67 (dt, J = 12.9, 6.4 Hz, 1H), 2.58–2.49 (m, 2H), 2.16 (s, 3H), 1.93 (ddd, J = 13.2, 6.0, 4.4 Hz, 1H), 1.59 (ddd, J = 13.2, 7.6, 5.6 Hz, 1H), 1.24 (m, 6H), 1.17 (d, J = 6.8 Hz, 3H), 1.11-1.06 (m, 36H), 0.94 (s, 3H), 0.89 (s, 9H), 0.85 (s, 3H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ: 212.0, 170.8, 170.5, 155.9, 151.3, 136.6, 135.5, 128.4, 128.2, 128.1, 120.5, 119.7, 107.3, 82.6, 76.4, 72.8, 71.4, 69.9, 66.6, 58.5, 53.1, 52.2, 42.4, 37.8, 36.0, 29.9, 25.8, 24.6, 18.0, 18.0, 17.9, 17.2, 13.2, 13.1, 12.3, 11.3, -4.4, -5.0 ; MS (FAB) calculated for C55H93O10Si3 [M -CH₃O]⁺: 997.6077, found 997.6046.

(5)-Benzyl 2-((25,4R,6R)-Tetrahydro-6-((25,35,4R)-2,4-bishydroxy-3-methyl-5-(6-methyl-2-methyloxycarbonyl-3,5bistriisopropylsilyloxyphenyl)-pentyl)-5,5-dimethyl-4-*tert*butyldimethylsilyloxy-2*H*-pyran-2-yl)-2-methoxyacetate (+)-60. To a solution of THF/MeOH (3.5 mL/5.2 mL) was added Et₃B (1.0 M solution in hexane, 2.62 mL, 2.62 mmol). After stirring for 1 h, the reaction was cooled to -70 °C and aldol (+)-35 (539 mg, 0.524 mmol) in THF (7.0 mL) was added. After stirring for 30 min at -70 °C, NaBH₄ (198 mg, 5.24 mmol) was added, and the reaction was stirred for 16 h. The reaction was quenched with a mixture of MeOH/sat. NH₄Cl/30% aq. H₂O₂ (15 mL/15 mL/15 mL), and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification

by column chromatography (EtOAc/Hexane 1/12-1/8) gave diol (+)-60 (503 mg, 0.488 mmol, 93%, dr > 20:1) as a white foam.

(+)-**60**: $[\alpha]_{D}^{22}$ +14.8 (c 2.42, CHCl₃); IR (neat): 3490, 2947, 2867, 1732, 1589, 1468, 1258, 1165, 1100, 1069 cm⁻¹; ¹H NMR (400 MHz, $CDCl_3$) δ : 7.38–7.29 (m, 5H), 6.25 (s, 1H), 5.29 (d, J = 12.4 Hz, 1H), 5.18 (d, J = 12.0 Hz, 1H), 4.19 (dt, J = 9.6, 4,8 Hz, 1H), 4.01-3.99 (m, 2H), 3.87-3.83 (m, 4H), 3.73 (brs, 2H), 3.55-3.52 (m, 2H), 3.37 (s, 3H), 2.80 (ddd, J = 18.0, 14.0, 6.0 Hz, 2H), 2.19 (s, 3H), 1.92 (dt, J = 14.0, 4.4 Hz, 1H), 1.87–1.81 (dt, J = 14.4, 9.6 Hz, 1H), 1.65 (ddd, J = 14.4, 9.6, 5.6 Hz, 1H), 1.51 (dt, J = 6.8, 2.0 Hz, 1H), 1.45 (d, J = 14.4 Hz, 1H), 1.25 (sept, J = 7.6 Hz, 3H), 1.23 (sept, J = 7.6 Hz, 3H), 1.11-1.06 (m, 36H), 0.94 (d, J = 6.8 Hz, 3H), 0.89 (s, 9H), 0.85 (s,3H), 0.84 (s, 3H), 0.02 (s, 3H), 0.01 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) *b*: 170.6, 170.3, 155.6, 151.1, 137.1, 135.4, 128.5, 128.2, 120.7, 120.1, 107.1, 81.6, 81.5, 75.3, 72.2, 71.2, 66.9, 58.4, 52.1, 41.9, 38.9, 36.2, 33.5, 29.9, 25.8, 24.1, 18.1, 17.9, 15.5, 13.2, 13.1, 12.6, 6.1, -4.3, -4.9; MS (FAB) calculated for $C_{55}H_{95}O_{10}Si_3$ [M - CH₃O]⁺: 999.6233. found 999.6224.

(S)-Benzyl 2-((2S,4R,6R)-Tetrahydro-6-((2S,3S)-3-((3R)-6,8bistriisopropylsilyloxy-5-methyl-3,4-dihydroisocoumarinyl)-3methyl-2-tert-butyldimethylsilyloxypropyl)-5,5-dimethyl-4tert-butyldimethylsilyloxy-2H-pyran-2-yl)-2-methoxyacetate (+)-36. To a solution of diol (+)-60 (98.0 mg, 0.0951 mmol) in CH₂Cl₂ (1.0 mL) was added CSA (2.21 mg, 0.00951 mmol) at rt. After stirring for 2 h, the reaction was quenched with sat. NaHCO₃ (1 mL) and the crude was extracted with EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded the crude lactone. To a solution of crude lactone in CH₂Cl₂ (0.5 mL) was added 2,6-lutidine (0.0665 mL, 0.571 mmol), followed by TBSOTf (0.0655 mL, 0.285 mmol), at 0 °C. After stirring for 2 h, the reaction was diluted with Et₂O (2 mL) and quenched with sat. NaHCO₃ (1 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/40-1/20) gave TBS-ether (+)-36 (82.1 mg, 0.0738 mmol, 78% over 2 steps) as a white foam.

(+)-36: $[\alpha]_D^{25}$ +48.9 (c 0.98, CHCl₃); IR (neat): 2947, 2892, 2867, 1727, 1591, 1568, 1472, 1353, 1249, 1172, 1068 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.38–7.29 (m, SH), 6.32 (s, 1H), 5.26 (d, *J* = 12.4 Hz, 1H), 5.12 (d, *J* = 12.0 Hz, 1H), 4.20–4.11 (m, 3H), 3.97 (d, *J* = 5.2 Hz, 1H), 3.62 (dd, *J* = 7.2, 4.4 Hz, 1H), 3.43 (d, *J* = 6.8 Hz, 1H), 3.39 (s, 3H), 3.08 (d, *J* = 16.0 Hz, 1H), 2.64 (dd, *J* = 16.4, 12.4 Hz, 1H), 1.58 (m, 1H), 1.36–1.24 (m, 6H), 1.14–1.08 (m, 39H), 0.93 (s, 3H), 0.89 (s, 9H), 0.84 (s, 3H), 0.78 (s, 6H), 0.05 (s, 6H), 0.03 (s, 3H), -0.04 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 170.2, 163.3, 158.4, 157.2, 141.3, 135.3, 128.4, 128.3, 128.2, 118.0, 110.4, 109.7, 83.3, 79.3, 77.1, 72.9, 69.8, 69.0, 66.6, 58.7, 39.9, 38.1, 33.5, 30.2, 25.9, 25.8, 25.0, 18.1, 18.0, 18.0, 16.9, 13.3, 11.9, 8.6, -3.4, -4.3, -4.9; MS (FAB) calculated for C₆₁H₁₀₉O₁₀Si₄ [M + H]⁺: 1113.7098, found 1113.7126.

(2R,4R,6S)-2-((2S,3S)-3-((3R)-6,8-Bistriisopropylsilyloxy-5methyl-3,4-dihydroisocoumarinyl)-3-methyl-2-tert-butyldimethylsilyloxypropyl)-3,3-dimethyl-6-((S)-methoxy-(N-(2-(trimethylsilyl)ethoxycarbonyl)amino)-methyl)-4-tert-butyldimethylsilyloxytetrahydropyran (+)-5. To a solution of TBSether (+)-36 (272 mg, 0.244 mmol) in MeOH (2.4 mL) was added 10% Pd-C (27.2 mg, 10% w/w) and hydrogenated (H_2 , 1 atm). After 1 h, the catalyst was filtered through a pad of Celite, followed by washing with EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded a crude carboxylic acid. To a solution of the crude carboxylic acid in THF (4.9 mL) was added NMM (0.0537 mL, 0.489 mmol), followed by EtOCOCl (0.0468 mL, 0.489 mmol), at -20 °C. After stirring for 20 min, NaN₃ (79.3 mg, 1.22 mmol) in H₂O (0.5 mL) was added to the reaction. After 20 min, NaHCO₃ (1 mL) was added and the mixture was extracted with EtOAc. The combined organic layers were washed with brine and dried over MgSO4, and toluene (4.9 mL) was added. Filtration and concentration afforded a toluene solution of the crude acyl azide. This solution was heated to 100 °C and stirred for 30 min. The reaction mixture was cooled to rt, and 2-(trimethylsilyl)ethanol

(0.350 mL, 2.44 mmol) was added. After stirring for 17 h at 100 $^{\circ}$ C, the reaction was cooled to rt and the solvent was evaporated. Purification by column chromatography (EtOAc/Hexane 1/20) gave Teoc-protected hemiaminal (+)-5 (262 mg, 0.230 mmol, 94% over 3 steps) as a white foam.

(+)-5: $[\alpha]_{D}^{21}$ +41.2 (c 0.66, CHCl₃); IR (neat): 2950, 2895, 2867, 1727, 1592, 1568, 1472, 1353, 1250, 1173, 1067 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.30 (s, 1H), 5.45 (d, J = 8.0 Hz, 1H), 4.80 (d, J = 8.0Hz, 1H), 4.21-4.10 (m, 4H), 4.00 (d, J = 6.4 Hz, 1H), 3.57 (brs, 1H), 3.36-3.34 (m, 4H), 3.03 (d, J = 12.8 Hz, 1H), 2.59 (dd, J = 12.8, 9.6 Hz, 1H), 2.27 (t, J = 10.8 Hz, 1H), 2.09 (s, 3H), 1.94 (t, J = 4.8 Hz, 1H), 1.81 (t, J = 8.8 Hz, 1H), 1.61 (dd, J = 18.0, 9.6 Hz, 1H), 1.46 (dt, I = 11.2, 4.8 Hz, 1H), 1.34-1.25 (m, 6H), 1.12-1.08 (m, 36H), 1.05 (d, J = 5.2 Hz, 3H), 0.96 (s, 3H), 0.89 (s, 9H), 0.87 (m, 2H), 0.85 (s, 3H), 0.85 (s, 3H), 0.87 (m, 2H), 0.85 (s, 3H), 0.85 (s, 33H), 0.77 (s, 9H), 0.07 (s, 3H), 0.05 (s, 3H), 0.04 (s, 3H), 0.00 (s, 9H), -0.04 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ: 163.4, 158.6, 157.4, 156.9, 141.3, 118.0, 110.5, 109.7, 84.2, 79.3, 77.4, 73.4, 68.6, 67.5, 63.4, 55.9, 39.5, 37.4, 32.5, 31.5, 29.9, 26.5, 26.0, 25.8, 20.7, 18.1, 18.0, 17.9, 17.6, 13.2, 13.1, 11.8, 8.7, -1.5, -3.5, -4.5, -4.9, -5.1; MS (FAB) calculated for $C_{56}H_{108}O_{10}NSi_5 [M - C_3H_7]^+$: 1094.6820, found 1094.6835

(+)-11,15-Bis-tert-butyldimethylsilyl-21,23-bistriisopropylsilyl-7-N-(2-(trimethylsilyl)ethoxycarbonyl)-5-(trimethylsilyl)ethoxymethylirciniastatin A (+)-38. To a solution of carboxylic acid (-)-6 (24.8 mg, 0.0814 mmol) in CH₂Cl₂ (0.41 mL) was added *i*-Pr₂NEt (0.0156 mL, 0.0895 mmol), followed by PivCl (0.0105 mL, 0.0854 mmol), at 0 $^\circ$ C. After stirring for 30 min, the reaction was quenched with sat. NaHCO3 (1 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. The unpurified mixed anhydride 37 was azeotroped with benzene $(3 \times 1 \text{ mL})$, placed on the vacuum pump for 30 min, and then dissolved in THF (0.7 mL). To a solution of a Teoc-protected hemiaminal (+)-5 (61.7 mg, 0.0542 mmol) in THF (1.1 mL) was added activated 4 Å molecular sieves (108 mg) and the solution of the mixed anhydride 37. The mixture was cooled to -78 °C, and LHMDS (1.6 M solution in THF, 0.136 mL, 0.217 mmol) was added. After stirring for 30 min, the reaction was allowed to warm to -60 °C and stirred for 6.5 h. Sat. NH₄Cl (2 mL) was added, and the reaction was allowed to warm to rt. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO4, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/40-1/20) gave amide (+)-38 (42.2 mg, 0.0297 mmol, 55%, 93% BRSM) as a colorless oil.

(+)-38: $[\alpha]_{D}^{21}$ +41.2 (c 0.66, CHCl₃); IR (neat): 2950, 2895, 2867, 1727, 1592, 1568, 1472, 1353, 1250, 1173, 1067 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.30 (s, 1H), 5.65 (d, J = 4.0 Hz, 1H), 5.15 (d, J = 3.6 Hz, 1H), 4.77 (d, J = 7.6 Hz, 1H), 4.67 (d, J = 5.2 Hz, 1H), 4.61 (d, J = 5.2 Hz, 1H), 4.32-4.29 (m, 3H), 4.17 (ddd, J = 9.2, 6.4, 1.2 Hz, 1H), 4.11 (dt, *J* = 8.0, 2.8 Hz, 1H), 3.65 (ddd, *J* = 7.2, 3.6, 2.0 Hz, 1H), 3.61-3.52 (m, 3H), 3.35 (s, 3H), 3.27 (s, 3H), 3.16 (d, J = 7.6 Hz, 1H), 3.06 (dd, *J* = 13.2, 1.2 Hz, 1H), 2.86 (dd, *J* = 13.2, 10.0 Hz, 1H), 2.30 (dd, J = 12.0, 7.6 Hz, 1H), 2.23–2.18 (m, 4H), 2.02 (m, 1H), 1.96 (m, 1H), 1.80 (m, 1H), 1.77 (s, 3H), 1.75-1.61 (m, 2H), 1.35-1.24 (m, 6H), 1.13-1.08 (m, 39H), 1.04 (m, 2H), 0.90 (s, 12H), 0.83 (m, 2H), 0.81 (s, 3H), 0.76 (s, 9H), 0.06-0.04 (s, 15H), -0.04 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ: 174.6, 163.5, 158.5, 157.3, 154.3, 142.7, 141.9, 118.3, 112.7, 110.5, 109.6, 95.1, 88.3, 81.0, 79.5, 75.4, 74.3, 72.7, 69.0, 66.0, 65.9, 58.0, 56.5, 40.3, 39.0, 38.8, 35.0, 30.3, 30.0, 29.7, 25.9, 25.8, 24.0, 22.8, 18.1, 18.0, 18.0, 17.6, 13.6, 13.3, 13.1, 12.0, 8.8, -1.5, -1.6, -3.3, -4.2, -4.9, -5.0; MS (FAB) calculated for $C_{70}H_{134}O_{14}NSi_6 [M - C_3H_7]^+$: 1380.8420, found 1380.8413.

(+)-Irciniastatin A/Psymberin [(+)-1]. To a solution of amide (+)-38 (13.3 mg, 9.32 μ mol) in DMF (0.9 mL) was added TASF (102 mg, 0.373 mmol), and the reaction was heated to 50 °C. After stirring for 16 h, the reaction was quenched with sat. NH₄Cl (1 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/1–2/1) gave (+)-irciniastatin A/psymberin [(+)-1] (3.02 mg, 4.96 μ mol, 53%) as a white solid.

(+)-1: $[\alpha]_D^{25}$ +26.6 (c 0.11, CHCl₃); IR (neat): 3378, 2927, 1656, 1619, 1519, 1462, 1376, 1253, 1173, 1106, 1069 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 11.1 (s, 1H), 7.10 (d, *J* = 8.0 Hz, 1H), 5.45 (dd, *J* = 8.0, 7.2 Hz, 1H), 4.80 (s, 2H), 4.53 (dt, *J* = 9.2, 3.2 Hz, 1H), 4.41 (s, 1H), 4.35 (s, 1H), 4.30 (m, 1H), 3.94 (d, *J* = 8.4 Hz, 1H), 3.88 (m, 1H), 3.74 (m, 1H), 3.67 (dd, *J* = 8.4, 4.8 Hz, 1H), 3.54 (dd, *J* = 13.2, 1H), 3.38 (s, 6H), 2.88 (dd, *J* = 13.2, 2.8 Hz, 1H), 2.81 (dd, *J* = 13.2, 10.0 Hz, 1H), 2.03 (s, 3H), 1.88–1.78 (m, 2H), 1.75 (s, 3H), 1.63 (m, 1H), 1.10 (d, *J* = 6.0 Hz, 3H), 0.97 (s, 3H), 0.92 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 176.4, 173.6, 170.5, 161.1, 142.0, 139.7, 113.2, 113.0, 101.6, 101.3, 81.9, 80.6, 79.6, 78.3, 73.9, 73.1, 73.1, 71.4, 57.9, 56.3, 42.7, 38.8, 37.6, 32.2, 29.7, 28.5, 23.1, 22.7, 13.6, 10.5, 9.2; MS (FAB) calculated for C₃₀H₄₄O₁₀N [M - CH₃O]⁺: 578.2965, found 578.2967.

(+)-5,11,21,23-Tetrakis-*tert*-butyldimethylsilylirciniastatin A (+)-61. To a solution of (+)-irciniastatin A [(+)-1] (10.9 mg, 0.0179 mmol) in CH₂Cl₂ (1.8 mL) was added 2,6-lutidine (29.2 μ L, 0.251 mmol) and TBSOTf (28.8 μ L, 0.125 mmol) at 0 °C. After stirring for 1 h, the reaction was quenched with sat. NaHCO₃ (0.6 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/20–1/10) gave TBS-ether (+)-61 (6.80 mg, 6.38 μ mol, 36%) as a colorless oil.

(+)-**61**: $[\alpha]_{21}^{21}$ +28.0 (c 0.05, CHCl₃); IR (neat): 2932, 2359, 1728, 1684, 1472, 1253, 1073 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.09 (d, *J* = 10.0 Hz, 1H), 6.28 (s, 1H), 5.28 (dd, *J* = 10.0, 5.5 Hz, 1H), 4.79 (s, 1H), 4.75 (s, 1H), 4.40 (d, *J* = 1.5 Hz, 1H), 4.26 (ddd, *J* = 11.5, 7.4, 2.6 Hz, 1H), 4.00–3.92 (m, 2H), 3.68 (ddd, *J* = 9.3, 3.4, 1.5 Hz, 1H), 3.63 (s, 1H), 3.54 (dd, *J* = 7.8, 3.4 Hz, 1H), 3.35 (s, 3H), 3.21 (s, 3H), 3.11 (dd, *J* = 16.3, 2.7 Hz, 1H), 2.77 (dd, *J* = 16.6, 11.7 Hz, 1H), 2.32 (dd, *J* = 13.6, 6.8, 4.0 Hz, 1H), 1.80 (dt, *J* = 7.2, 2.0 Hz, 1H), 1.73 (s, 3H), 1.60 (m, 2H), 1.10 (d, *J* = 6.8 Hz, 4H), 1.04–0.89 (m, 42H), 0.25–0.03 (m, 21H), -0.01 (s, 3H); MS (ESI) calculated for C₅₅H₁₀₃O₁₁NNaSi₄ [M + Na]⁺: 1088.6500, found 1088.6486.

(25,35)-N-((S)-((25,4R,6R)-6-((S)-3-((R)-6,8-Bis((tert-butyldimethylsilyl)oxy)-5-methyl-1-oxoisochroman-3-yl)-2-oxobutyl)-4-((tert-butyldimethylsilyl)oxy)-5,5-dimethyltetrahydro-2H-pyran-2-yl)(methoxy)methyl)-2-((tert-butyldimethylsilyl)oxy)-3-methoxy-5-methylhex-5-enamide (+)-40. To a solution of TBS-ether (+)-61 (7.00 mg, 6.60 μ mol) in CH₂Cl₂ (0.66 mL) were added PhI(OAc)₂ (7.5 mg, 0.0200 mmol) and AZADO (2.2 mg, 0.0100 mmol). After stirring for 7 h, the reaction was then diluted with CH₂Cl₂ (3 mL) and quenched with sat. Na₂S₂O₃ (2.5 mL) at 0 °C. After stirring for 30 min at rt, the mixture was extracted with EtOAc and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by preparative TLC (EtOAc/ Hexane 1/4) gave ketone (+)-40 (5.60 mg, 5.30 μ mol, 80%) as a colorless oil.

(+)-40: $[\alpha]_{D}^{21}$ +44.3 (c 0.22, CHCl₃); IR (neat): 3169, 2929, 2863, 2360, 1741, 1685, 1589, 1496, 1462, 1251, 1068 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.14 (d, J = 10.0 Hz, 1H, N-7), 6.29 (s, 1H, C-22), 5.16 (dd, J = 10.4, 3.6 Hz, 1H, C-8), 4.78 (d, J = 13.2 Hz, 2H, C-1), 4.50-4.44 (m, 2H, C-5, C-17), 4.08 (dd, J = 7.3, 4.3 Hz, 1H, C-13), 3.94 (m, 1H, C-9), 3.75 (ddd, J = 9.2, 4.0, 2.0 Hz, 1H, C-4), 3.62 (dd, J = 4.8, 2.4 Hz, 1H, C-11), 3.39 (s, 3H), 3.33 (s, 3H), 3.04 (dd, J = 17.4, 7.7 Hz, 1H), 2.96–2.88 (m, 3H, C-16, C-18), 2.55 (dd, J = 16.5, 11.6, 1H, C-18), 2.33 (dd, J = 14.5, 9.1 Hz, 1H, C-3), 2.15 (dd, J = 15.0, 3.4 Hz, 1H, C-3), 2.00 (s, 3H), 1.80 (m, 1H), 1.74 (s, 3H), 1.58-1.49 (m, 1H), 1.31 (d, J = 6.8 Hz, 4H), 1.26 (s, 2H), 1.03–0.85 (m, 42H), 0.25-0.22 (m, 12H), 0.15 (s, 3H), 0.07-0.05 (m, 9H); ¹³C NMR (100 MHz, CDCl₃) δ: 209.7, 172.5, 142.6, 140.7, 118.8, 112.6, 110.6, 110.1, 82.4, 81.3, 76.0, 74.4, 73.2, 58.0, 56.1, 50.8, 43.0, 37.9, 37.5, 31.0, 30.0, 29.7, 26.0, 25.9, 25.8, 25.7, 22.8, 18.5, 18.3, 18.2, 18.0, 13.3, 11.7, -4.2, -4.2, -4.3, -4.4, -4.5, -4.9, -5.2; MS (ESI) calculated for $C_{55}H_{101}O_{11}NNaSi_4 [M + Na]^+$: 1086.6344, found 1086.6335.

(2R,4R,6S)-2-Allyl-6-((R)-2-hydroxy-1-methoxyethyl)-3,3dimethyltetrahydro-2*H*-pyran-4-ol (+)-42. To a solution of BOMether (+)-23 (203 mg, 0.557 mmol) in CH₃CN/H₂O (5.6 mL/0.3 mL) was added LiBF₄ (783 mg, 8.35 mmol) at rt. The reaction mixture was stirred at reflux for 3 h. The mixture was quenched with sat. NaHCO₃ (15 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/4–1/1) gave diol (+)-42 (123 mg, 0.501 mmol, 90%) as a white solid.

(+)-42: $[\alpha]_{D}^{22}$ +12.3 (c 0.71, CHCl₃); IR (neat): 3734, 3388, 2148, 1683, 1558, 1091 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 5.83 (ddt, *J* = 16.8, 10.0, 7.2 Hz, 1H), 5.08 (dd, *J* = 16.8, 10.0 Hz, 2H), 3.92 (ddd, *J* = 8.7, 4.2, 4.0 Hz, 1H), 3.77 (dd, *J* = 12.0, 4.2 Hz, 1H), 3.69 (dd, *J* = 12.0, 4.0 Hz, 1H), 3.62 (dd, *J* = 9.1, 4.4 Hz, 1H), 3.45 (s, 3H), 3.35 (ddd, *J* = 8.7, 4.4, 4.4 Hz, 1H), 3.27 (dd, *J* = 10.8, 2.5 Hz, 1H), 2.43 (m, 1H), 2.22 (m, 2H), 2.00 (ddd, *J* = 13.6, 4.4, 4.4 Hz, 1H), 1.79 (ddd, *J* = 13.6, 9.1, 4.4 Hz, 1H), 1.64 (brs, 1H) 1.25 (s, 3H), 0.92 (s, 3H); ¹³C NMR (100 MHz, CD₃OD) δ : 138.2, 116.5, 81.4, 80.7, 61.8, 58.9, 39.6, 34.9, 30.4, 28.2, 25.9, 23.9 ; MS (EI) calculated for C₁₀H₁₉O₄ [M - C₃H₅]⁺: 203.1283, found 203.1277.

(2*R*,4*R*,6*S*)-2-Allyl-4-(benzyloxy)-6-((*R*)-2-(benzyloxy)-1methoxyethyl)-3,3-dimethyltetrahydro-2*H*-pyran (–)-43. To a solution of diol (+)-42 (14.7 mg, 0.0602 mmol) in THF (0.6 mL) was added NaH (12.0 mg, 0.301 mmol), followed by TBAI (2.2 mg, 6.02 μ mol) and BnBr (0.0357 mL, 0.301 mmol), at rt. The reaction mixture was stirred at reflux for 2 h. The mixture was quenched with sat. NH₄Cl (2 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/40–1/20) gave Bn-ether (–)-43 (20.4 mg, 0.0479 mmol, 80%) as a colorless oil.

(-)-43: $[\alpha]_{D}^{22}$ -0.98 (c 0.51, CHCl₃); IR (neat): 2863, 1639, 1454, 1094 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.34–7.28 (m, 10H), 5.74 (ddt, *J* = 17.2, 10.4, 6.8 Hz, 2H), 4.97 (m, 2H), 4.64 (d, *J* = 12.4 Hz, 1H), 4.57 (d, *J* = 12.2 Hz, 1H), 4.50 (d, *J* = 12.2 Hz, 1H), 4.41 (d, *J* = 12.4 Hz, 1H), 3.89 (m, 1H), 3.75 (m, 1H), 3.48–3.45 (m, 2H), 3.45 (s, 3H), 3.27 (dd, *J* = 9.0, 4.0 Hz, 1H), 3.21 (dd, *J* = 10.6, 2.6 Hz, 1H) 2.38 (m, 1H), 2.18 (dd, *J* = 13.6, 6.8 Hz, 1H), 2.07 (dt, *J* = 14.0, 4.0 Hz, 1H), 1.78 (m, 1H), 0.99 (s, 3H), 0.94 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 137.0, 128.3, 128.3, 128.2, 127.6, 127.6, 127.5, 127.5, 127.4, 116.0, 87.7, 79.7, 79.1, 73.5, 71.1, 70.4, 70.3, 58.8, 38.3, 33.6, 25.5, 25.5 ; MS (EI) calculated for C₃₈H₄₀O₆ [M]⁺: 556.2825, found 556.2840.

1-((2R,4R,6S)-4-(Benzyloxy)-6-((R)-2-(benzyloxy)-1-methoxyethyl)-3,3-dimethyltetrahydro-2H-pyran-2-yl)butan-2-one (+)-44. To a solution of Bn-ether (-)-43 (117 mg, 0.276 mmol) in THF/H₂O (2.8/0.3 mL) was added NMO (48.6 mg, 0.415 mmol), followed by OsO_4 (0.196 M solution in THF, 0.028 mL, 5.53 μ mol), at 0 °C. The reaction mixture was stirred at rt for 12 h. The mixture was quenched with sat. Na₂S₂O₃ (3 mL) and stirred for 1 h at rt. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine and dried over MgSO4. Filtration and concentration afforded a crude diol. To a solution of the diol in MeOH/H₂O (5.0 mL/0.6 mL) was added NaIO₄ (88.7 mg, 0.415 mmol), and the mixture was stirred at rt. After 1 h, EtOAc and H₂O were added. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine and dried over MgSO4. Filtration and concentration afforded a crude aldehyde. To a solution of EtMgBr (1.0 M solution in THF, 1.66 mL, 1.66 mmol) was added the crude aldehyde dropwise at 0 °C. After stirring for 1 h, the reaction was quenched with sat. NH₄Cl (6 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded a crude ethyl alcohol. To a solution of the ethyl alcohol in CH₂Cl₂ (0.6 mL) was added PhI(OAc)₂ (134 mg, 0.415 mmol), followed by 1-Me-AZADO (4.6 mg, 0.0276 mmol), at rt. After stirring for 3 h, the reaction was then diluted with Et₂O (3 mL) and quenched with sat. Na₂S₂O₃ (1.5 mL) at 0 $^\circ\text{C}.$ After stirring for 1 h at rt, the mixture was extracted with EtOAc and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/8) gave ketone (+)-44 (102 mg, 0.224 mmol, 81% over 4 steps) as a colorless oil.

(+)-44: $[\alpha]_D^{27}$ +2.3 (c 0.61, CHCl₃); IR (neat): 2933, 2872, 2359, 1714, 1453, 1363, 1094 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.36–7.27 (m, 10H), 4.65 (d, *J* = 12.0 Hz, 1H), 4.53 (s, 2H), 4.41 (d, *J* = 11.6 Hz, 1H), 3.94 (m, 1H), 3.84 (m, 1H), 3.66 (d, *J* = 8.0 Hz, 1H), 3.47–3.43 (m, 2H), 3.43 (s, 3H), 3.31 (dd, *J* = 9.2, 4.0 Hz, 1H), 2.79 (dd, *J* = 15.6, 10.0 Hz, 1H), 2.41 (m, 3H), 2.08 (dt, *J* = 13.6, 4.4 Hz, 1H), 1.78 (m, 1H), 1.00 (t, *J* = 7.2 Hz, 3H), 0.97 (s, 3H), 0.93 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 210.5, 138.8, 138.6, 128.3, 128.3, 128.2, 127.6, 127.5, 127.5, 127.4, 79.3, 78.9, 76.0, 73.4, 71.0, 69.0, 58.4, 42.1, 37.8, 37.4, 25.4, 24.2, 7.6 ; MS (EI) calculated for C₂₈H₃₈O₅ [M]⁺: 454.2719, found 454.2698.

Methyl 2-((2R,3S)-5-((2R,4R,6S)-4-(Benzyloxy)-6-((R)-2-(benzyloxy)-1-methoxyethyl)-3,3-dimethyltetrahydro-2H-pyran-2yl)-2-hydroxy-3-methyl-4-oxopentyl)-3-methyl-4,6-bis((triisopropylsilyl)oxy)benzoate (+)-45. A solution of ketone (+)-44 (68.0 mg, 0.150 mmol) in CH_2Cl_2 (2.2 mL) was cooled to -78 °C, and PhBCl₂ (70.0 µL, 0.539 mmol) was added. After stirring for 4 min, i-Pr₂NEt (0.117 mL, 0.673 mmol) was added dropwise. After stirring for 1 h at -78 °C, aldehyde 32 (200.6 mg, 0.374 mmol) in CH₂Cl₂ (0.8 mL) was added to the boron enolate dropwise. After stirring for 1 h at -78 °C, the reaction was guenched with the mixture of MeOH/pH 7 buffer (5.0/5.0 mL). After 15 min at -78 °C, sat. NaHCO₃ aq. was added to neutralize the reaction mixture to pH 7, and the mixture was stirred for 1 h at 0 °C. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO4, and concentrated. Purification by column chromatography (EtOAc/ Hexane 1/15-1/8) gave aldol (+)-45 (121 mg, 0.122 mmol, 82%) as a white foam.

(+)-45: $[\alpha]_{D}^{25}$ +21.4 (c 0.69, CHCl₃); IR (neat): 2945, 2867, 1708, 1589, 1467, 1416, 1342, 1265, 1193, 1166, 1071, 919 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.37–7.24 (m, 10H), 6.26 (s, 1H), 4.65 (d, J = 12.2 Hz, 1H), 4.52 (s, 1H), 4.41 (d, J = 12.2 Hz, 1H), 4.03 (m, 1H), 3.91 (m, 1H), 3.87 (d, J = 8.4 Hz, 1H), 3.82 (s, 3H), 3.68 (dd, J = 10.8, 2.4 Hz, 1H), 3.53 (m, 2H), 3.47-3.40 (m, 1H), 3.42 (s, 3H), 3.31 (dd, J = 10.0, 4.0 Hz, 1H), 2.90 (dd, J = 16.4, 9.6 Hz, 1H), 2.81 (dd, I = 14.4, 3.2 Hz, 1H), 2.66 (t, I = 6.0 Hz, 1H), 2.53-2.44 (m, 1)2H), 2.15 (s, 3H), 2.15-2.10 (m, 1H), 1.72 (m, 1H), 1.23 (sept, J = 7.6 Hz, 6H), 1.14–1.06 (m, 39H), 0.97 (s, 3H), 0.94 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ: 212.5, 171.1, 156.1, 151.6, 138.7, 138.6, 136.6, 128.3, 128.3, 128.2, 127.6, 127.5, 127.5, 127.3, 120.6, 119.8, 107.4, 78.8, 77.3, 77.2, 77.0, 76.7, 75.3, 73.4, 71.4, 70.9, 69.2, 58.5, 53.5, 52.3, 42.2, 37.8, 35.9, 25.3, 23.9, 18.0, 17.9, 13.2, 13.0, 12.3, 11.1; MS (FAB) calculated for $C_{56}H_{87}O_9Si_2$ [M - CH₃O]⁺: 959.5889, found 959.5894

Methyl 2-((2R,3S,4S)-5-((2R,4R,6S)-4-(Benzyloxy)-6-((R)-2-(benzyloxy)-1-methoxyethyl)-3,3-dimethyltetrahydro-2Hpyran-2-yl)-2,4-dihydroxy-3-methylpentyl)-3-methyl-4,6-bis-((triisopropylsilyl)oxy)benzoate (+)-62. To a solution of aldol (+)-45 (296 mg, 0.299 mmol) in THF/MeOH (3.3 mL/1.2 mL) cooled to -78 °C was added Et₂BOMe (1.0 M solution in THF, 0.60 mL, 0.598 mmol). After stirring for 1 h, to the reaction was added NaBH₄ (56.5 mg, 1.49 mmol). After 1 h, the reaction mixture was warmed to 0 °C. After 1.5 h, the reaction was quenched with a mixture of EtOAc/H2O (4.0/4.0 mL) and MeOH/30% aq. H2O2 (6.5/6.5 mL) and stirred for 1 h. The mixture was extracted with EtOAc, and the combined organic layers were treated with solid Na2S2O3 to destroy any remaining peroxide. The organic layer was then filtered and washed with sat. Na₂S₂O₃, dried over MgSO₄, and concentrated. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/8-1/6) gave diol (+)-62 (231 mg, 0.232 mmol, 78%) as a white foam.

(+)-62: $[\alpha]_D^{26}$ +5.9 (c 0.21, CHCl₃); IR (neat): 3471, 2944, 2866, 1727, 1588, 1466, 1341, 1259, 1164, 1097 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 7.33–7.26 (m, 10H), 6.26 (s, 1H), 4.63 (d, *J* = 12.4 Hz, 1H), 4.58 (d, *J* = 12.4 Hz, 1H), 4.50 (d, *J* = 12.4 Hz, 1H), 4.58 (d, *J* = 12.4 Hz, 1H), 4.50 (d, *J* = 12.4 Hz, 1H), 4.41 (d, *J* = 12.4 Hz, 1H), 4.10–4.06 (m, 1H), 3.95 (s, 2H), 3.82 (s, 3H), 3.70 (dt, *J* = 10.4, 2.8 Hz, 1H), 3.54 (dd, *J* = 10.8, 4.0 Hz, 1H), 3.47–3.40 (m, 1H), 3.40 (s, 3H), 3.26 (dd, *J* = 8.8, 3.2 Hz, 1H), 2.78 (d, *J* = 6.4 Hz, 2H), 2.17 (s, 3H), 2.08–2.05 (m, 1H), 1.95–1.91 (m, 1H), 1.73–1.66

(m, 1H), 1.59 (s, 2H), 1.48–1.43 (m, 2H), 1.26 (sept, *J* = 7.6 Hz, 6H), 1.11–1.06 (m, 36H), 0.98–0.93 (m, 9H); ¹³C NMR (100 MHz, CDCl₃) δ : 162.7, 158.7, 157.4, 140.8, 139.0, 138.3, 128.3, 128.2, 127.5, 127.5, 127.3, 127.3, 117.9, 109.9, 109.3, 79.7, 78.2, 77.3, 77.2, 77.0, 76.7, 76.5, 73.6, 71.1, 70.6, 69.8, 59.1, 43.8, 37.7, 29.8, 25.9, 25.2, 25.0, 18.1, 18.0, 18.0, 13.3, 13.1, 11.7, 10.4, -4.1, -4.7; MS (FAB) calculated for C₅₇H₉₂O₁₀Si₂ [M - H]⁺: 991.6045, found 991.6082.

(R)-3-((2S,3S)-4-((2R,4R,6S)-4-(Benzyloxy)-6-((R)-2-(benzyloxy)-1-methoxyethyl)-3,3-dimethyltetrahydro-2H-pyran-2-yl)-3-((tert-butyldimethylsilyl)oxy)butan-2-yl)-5-methyl-6,8-bis-((triisopropylsilyl)oxy)isochroman-1-one (+)-46. To a solution of diol (+)-62 (254 mg, 0.255 mmol) in $\rm CH_2Cl_2$ (5.1 mL) was added CSA (6.0 mg, 0.0255 mmol) at 0 °C. After stirring for 30 min, the reaction was quenched with sat. NaHCO3 (1.5 mL) and the crude was extracted with EtOAc. The combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded the crude lactone. To a solution of crude lactone in CH_2Cl_2 (5.1 mL) was added 2,6-lutidine (0.12 mL, 1.022 mmol), followed by TBSOTf (0.12 mL, 0.511 mmol), at 0 °C. After stirring for 1 h, the reaction was diluted with Et₂O (2 mL) and quenched with sat. NaHCO₃ (2 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/20-1/15) gave TBS-ether (+)-46 (213 mg, 0.198 mmol, 78% over 2 steps) as a white foam.

(+)-46: $[\alpha]_{D}^{27}$ +36.3 (c 0.14, CHCl₃); IR (neat): 2945, 2866, 1725, 1590, 1567, 1471, 1351, 1244, 1169, 1068 cm⁻¹; ¹H NMR (400 MHz, $CDCl_3$) δ : 7.42–7.28 (m, 10H), 6.38 (s, 1H), 4.67 (d, J = 12.0 Hz, 1H), 4.62 (d, J = 12.0 Hz, 1H), 4.58 (d, J = 12.0 Hz, 1H), 4.45 (d, J = 12.0 Hz, 1H), 4.24 (t, J = 9.2 Hz, 1H), 4.17–4.14 (m, 1H), 4.05–4.03 (m, 1H), 3.76 (dd, J = 10.4, 4.0 Hz, 1H), 3.64 (dd, J = 10.0, 5.2 Hz, 1H), 3.53-3.51 (m, 1H), 3.53 (s, 3H), 3.42-3.36 (m, 2H), 3.06 (d, J = 16.4 Hz, 1H), 2.58 (dd, J = 16.0, 12.0 Hz, 1H), 2.30–2.20 (m, 1H), 2.12 (s, 3H), 2.08–2.00 (m, 2H), 1.84–1.80 (m, 1H), 1.67 (t, J = 11.2 Hz, 1H), 1.37 (sept, I = 7.6 Hz, 6H), 1.20–1.11 (m, 39H), 1.07 (s, 3H), 1.01 (s, 3H), 0.81 (s, 9H), 0.00 (s, 6H); ¹³C NMR (100 MHz, CDCl₃) *δ*: 163.3, 158.7, 157.5, 141.1, 138.8, 138.2, 128.3, 128.3, 128.2, 127.7, 127.5, 127.4, 117.9, 110.4, 109.8, 81.5, 80.1, 79.2, 73.4, 71.7, 69.8, 68.9, 58.9, 39.7, 37.6, 32.7, 29.9, 25.9, 25.7, 25.6, 18.0, 17.9, 13.2, 13.1, 13.0, 11.9, 8.7, -3.6, -5.0 ; MS (FAB) calculated for $C_{59}H_{95}O_9Si_3$ [M - C_3H_7]⁺: 1031.6284, found 1031.6305

(*R*)-3-((2*S*,3*S*)-3-((*tert*-Butyldimethylsilyl)oxy)-4-((2*R*,4*R*,6*S*)-4-hydroxy-6-((*R*)-2-hydroxy-1-methoxyethyl)-3,3-dimethyltetra-hydro-2*H*-pyran-2-yl)butan-2-yl)-5-methyl-6,8-bis((*tri-isopropylsilyl*)oxy)isochroman-1-one (+)-47. To a solution of TBS-ether (+)-46 (73.9 mg, 0.0687 mmol) in MeOH (1.4 mL) was added 10% Pd-C (7.4 mg, 10% w/w) and hydrogenated (H₂, 1 atm). After 5 h, the catalyst was filtered through a pad of Celite, followed by washing with EtOAc and concentrated. Purification by column chromatography (EtOAc/Hexane 1/4–1/2) gave diol (+)-47 (58.4 mg, 0.0653 mmol, 95%) as a white foam.

(+)-47: $[\alpha]_D^{27}$ +58.3 (c 0.53, CHCl₃); IR (neat): 3440, 2947, 2868, 1705, 1591, 1567, 1472, 1411, 1386, 1353, 1248, 1172, 1085 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.28 (s, 1H), 4.27–4.19 (m, 2H), 4.04 (d, *J* = 12.0 Hz, 1H), 3.83 (dt, *J* = 9.2, 4.8 Hz, 1H), 3.63–3.61 (m, 2H), 3.53 (s, 3H), 3.45 (ddd, *J* = 12.0, 6.0, 2.6 Hz, 1H), 3.37 (brs, 1H), 3.21 (d, *J* = 10.4 Hz, 1H), 2.96 (dd, *J* = 16.4, 2.0 Hz, 1H), 2.74 (dd, *J* = 16.8, 12.8 Hz, 1H), 2.08 (s, 3H), 2.02–1.91 (m, 3H), 1.77–1.65 (m, 2H), 1.48 (brs, 1H), 1.29 (sept, *J* = 7.2 Hz, 6H), 1.11–1.07 (m, 36H), 1.04 (s, 3H), 0.95 (s, 3H), 0.88 (s, 3H), 0.83 (s, 9H), 0.08 (s, 3H), 0.00 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 164.1, 159.0, 157.7, 141.6, 118.1, 109.7, 109.5, 80.8, 79.4, 75.8, 72.5, 67.9, 62.2, 59.0, 39.8, 38.0, 34.5, 30.4, 29.1, 25.9, 25.8, 24.1, 18.0, 18.0, 17.9, 13.3, 13.1, 11.7, 8.6, -3.2, -4.7; MS (ESI) calculated for C₄₈H₉₁O₉Si₃ [M + H]⁺: 895.5965, found 895.5941.

(S)-2-((25,4R,6R)-6-((25,3S)-2-((tert-Butyldimethylsilyl)oxy)-3-((R)-5-methyl-1-oxo-6,8-bis((triisopropylsilyl)oxy)isochroman-3-yl)butyl)-4-hydroxy-5,5-dimethyltetrahydro-2*H*-pyran-2-yl)-2-methoxyacetic Acid (+)-48. To a solution of the diol (+)-47 (17.6 mg, 0.0196 mmol) in CH₃CN/pH 7 buffer (0.4/0.2 mL) were added NaClO₂ (80%) (6.7 mg, 0.0588 mmol) and DMN-

AZADO⁺BF₄⁻ (1.0 mg, 3.92 μ mol). After 4.5 h, 2-methyl-2-butene (2 mL) and sat. NH₄Cl (1 mL) were added. The mixture was extracted with CHCl₃, and the combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/2, then CHCl₃/MeOH 10%) gave carboxylic acid (+)-**48** (14.4 mg, 0.0158 mmol, 81%) as a white foam.

(+)-48: $[\alpha]_{28}^{28}$ +62.8 (c 0.96, CHCl₃); IR (neat): 3404, 2947, 2868, 1727, 1704, 1591, 1567, 1472, 1411, 1386, 1353, 1173, 1090 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.29 (s, 1H), 4.45 (dd, *J* = 11.0, 3.8 Hz, 1H), 4.25 (dq, *J* = 13.0, 2.8 Hz, 1H), 4.19–4.14 (m, 1H), 3.77 (d, *J* = 8.4 Hz, 1H), 3.70 (dd, *J* = 5.6, 3.2 Hz, 1H), 3.46–3.42 (m, 1H), 3.42 (s, 3H), 2.97 (dd, *J* = 16.8, 13.2 Hz, 1H), 2.85 (dd, *J* = 16.6, 2.6 Hz, 1H), 2.42 (t, *J* = 11.2 Hz, 1H), 2.17 (t, *J* = 6.6 Hz, 1H), 2.07 (s, 3H), 2.00–1.94 (m, 1H), 1.87–1.84 (m, 1H), 1.55 (dt, *J* = 10.8, 3.2 Hz, 1H), 1.30 (sept, *J* = 8.0 Hz, 6H), 1.34–1.26 (m, 1H), 1.13–1.08 (m, 36H), 1.03 (s, 3H), 0.97 (d, *J* = 7.2 Hz, 3H), 0.93 (s, 3H), 0.88 (s, 9H), 0.15 (s, 3H), 0.04 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ : 172.4, 166.2, 159.3, 157.8, 142.2, 118.1, 109.2, 109.2, 83.1, 79.8, 72.6, 68.1, 66.4, 58.2, 37.2, 36.9, 33.4, 31.7, 28.2, 26.1, 25.8, 18.0, 18.0, 17.9, 13.2, 13.1, 11.7, 8.1, -3.1, -4.5 ; MS (ESI) calculated for C₄₈H₈₉O₁₀Si₃ [M + H]⁺: 909.5764, found 909.5765.

2-(Trimethylsilyl)ethyl ((S)-((2S,4R,6R)-6-((2S,3S)-2-((tert-Butyldimethylsilyl)oxy)-3-((R)-5-methyl-1-oxo-6,8-bis((triisopropylsilyl)oxy)isochroman-3-yl)butyl)-4-hydroxy-5,5dimethyltetrahydro-2H-pyran-2-yl)(methoxy)methyl)carbamate (+)-49. To a solution of carboxylic acid (+)-47 (8.6 mg, 9.46 μmol) in THF (0.4 mL) was added NMM (3.1 μL, 0.028 mmol), followed by EtOCOCl (2.7 μ L, 0.028 mmol), at -20 °C. After stirring for 20 min, NaN₃ (3.7 mg, 0.057 mmol) in H_2O (0.023 mL) was added to the reaction. After 30 min, sat. NaHCO₃ (0.5 mL) was added and the mixture was extracted with EtOAc. The combined organic layers were washed with brine and dried over $MgSO_4$, and toluene (0.4) mL) was added. Filtration and concentration afforded a toluene solution of the crude acyl azide. This solution was heated to 100 °C and stirred for 1 h. The reaction mixture was cooled to rt, and 2-(trimethylsilyl)ethanol (0.014 mL, 0.095 mmol) was added. After stirring for 2.5 h at 100 °C, the reaction was cooled to rt and the solvent was evaporated. Purification by column chromatography (EtOAc/Hexane 1/8-1/4) gave Teoc-protected hemiaminal (+)-49 (8.3 mg, 8.13 μ mol, 86% over 2 steps) as a white foam.

(+)-49: $[\alpha]_D^{26}$ +111.4 (c 0.51, CHCl₃); IR (neat): 3447, 2947, 2867, 2360, 1707, 1590, 1567, 1472, 1352, 1248, 1173, 1070 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) & 6.30 (s, 1H), 5.42 (d, *J* = 9.2 Hz, 1H), 4.86 (d, *J* = 7.6 Hz, 1H), 4.21–4.17 (m, 3H), 4.11 (t, *J* = 7.0 Hz, 1H), 4.01 (brs, 1H), 3.66 (brs, 1H), 3.40–3.37 (m, 1H), 3.37 (s, 3H), 3.01 (d, *J* = 16.0 Hz, 1H), 2.62 (dd, *J* = 16.8, 12.0 Hz, 1H), 2.35–2.27 (m, 1H), 2.11 (s, 3H), 1.98–1.87 (m, 2H), 1.30 (sept, *J* = 7.6 Hz, 6H), 1.12–1.05 (m, 42H), 1.00 (s, 3H), 0.91 (s, 6H), 0.79 (s, 9H), 0.01 (m, 1SH); ¹³C NMR (100 MHz, CDCl₃) & 163.5, 158.6, 157.4, 141.4, 118.0, 110.4, 109.7, 84.1, 79.3, 79.3, 72.6, 68.6, 68.3, 63.4, 55.9, 39.6, 37.2, 32.8, 30.7, 29.8, 25.8, 25.7, 25.6, 18.0, 17.6, 17.5, 14.2, 13.3, 13.2, 13.2, 13.1, 11.9, 8.8, -1.6, -3.4, -5.1; MS (ESI) calculated for C₅₃H₁₀₁NO₁₀NaSi₄ [M + Na]⁺: 1046.6395, found 1046.6373.

2-(Trimethylsilyl)ethyl ((*S*)-((*2S*, *4R*, *6R*)-6-((*2S*, *3S*)-2-((*tert*-Butyldimethylsilyl)oxy)-3-((*R*)-5-methyl-1-oxo-6, *8*-bis-((triisopropylsilyl)oxy)isochroman-3-yl)butyl)-5,5-dimethyl-4-((triethylsilyl)oxy)tetrahydro-2*H*-pyran-2-yl) (methoxy)-methyl)((*2S*, *3S*)-3-methoxy-5-methyl-2-((*2*-(trimethylsilyl)ethoxy)methoxy)hex-5-enoyl)carbamate (+)-51. To a solution of hemiaminal (+)-49 (33.6 mg, 0.0328 mmol) in CH₂Cl₂ (0.8 mL) was added 2,6-lutidine (0.019 mL, 0.163 mmol), followed by TESOTf (0.019 mL, 0.082 mmol), at 0 °C. After stirring for 2 h, the reaction was quenched with sat. NaHCO₃ (1 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/40–1/20) gave give TES-ether **50** with unknown byproducts, which was taken on to the next step without further purification.

-78 °C, and LHMDS (1.0 M solution in THF, 0.13 mL, 0.131 mmol) was added. After stirring for 20 min, the solution of the mixed anhydride 37 was added. After stirring for 30 min, the reaction was allowed to warm to -60 °C and stirred for 1.5 h. Sat. NH₄Cl (2 mL) was added, and the reaction was allowed to warm to rt. The mixture was extracted with EtOAc, and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/40–1/20) gave amide (+)-**51** (21.8 mg, 0.0153 mmol, 47%, 91% BRSM) as a white foam.

(+)-51: $[\alpha]_{D}^{26}$ +81.2 (c 0.65, CHCl₃); IR (neat): 2951, 2868, 1727, 1590, 1568, 1471, 1411, 1351, 1248, 1172, 1066 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ : 6.34 (s, 1H), 5.68 (d, J = 4.4 Hz, 1H), 5.19 (d, J = 4.4 Hz, 1H), 4.81 (d, J = 8.0 Hz, 1H), 4.70 (d, J = 6.4 Hz, 1H), 4.65 (d, J = 6.4 Hz, 1H), 4.36-4.32 (m, 3H), 4.19-4.15 (m, 2H), 3.69-3.67 (m, 1H), 3.65-3.60 (m, 3H), 3.38 (s, 3H), 3.31 (s, 3H), 3.20 (d, J = 9.6 Hz, 1H), 3.11 (dd, J = 17.2 Hz, 1H), 2.90 (dd, J = 17.2, 12.8 Hz, 1H), 2.34-2.26 (m, 2H), 2.22 (s, 3H), 2.08-1.98 (m, 3H), 1.81-1.73 (m, 7H), 1.33 (sept, J = 7.6 Hz, 6H), 1.17–0.96 (m, 45H), 0.90 (s, 3H), 0.88 (s, 3H), 0.83 (s, 9H), 0.64 (q, J = 8.0 Hz, 6H), 0.12-0.03 (m, 30H); ¹³C NMR (100 MHz, CDCl₃) δ : 174.5, 158.5, 157.3, 154.3, 142.7, 141.9, 118.3, 112.6, 110.5, 109.6, 95.0, 81.0, 79.6, 76.7, 75.5, 72.7, 69.0, 66.0, 65.9, 58.1, 56.5, 40.3, 39.0, 38.7, 25.8, 24.0, 22.8, 18.0, 18.0, 17.6, 13.3, 13.1, 12.0, 8.7, 6.9, 5.1, -1.5, -1.6, - 3.3, -4.9; MS (ESI) calculated for C73H141NO14Si6Na [M + Na]+: 1446.8860, found 1446.8835.

(+)-15-tert-Butyldimethylsilyl-21,23-bistriisopropylsilyl-7-N-(2-(trimethylsilyl)ethoxycarbonyl)-5-(trimethylsilyl)ethoxymethylirciniastatin B (+)-52. To a solution of amide (+)-51 (9.5 mg, 6.67 μ mol) in THF (0.3 mL) was added 1 M HCl (0.020 mL, 0.020 mmol) at 0 °C. After stirring for 3 h at rt, the reaction was quenched with sat. NaHCO₃ (1 mL). The mixture was extracted with EtOAc, and the combined organic layers were washed with brine and dried over MgSO₄. Filtration and concentration afforded a crude alcohol. To a solution of the alcohol in CH₂Cl₂ (0.3 mL) were added PhI(OAc)₂ (6.5 mg, 0.020 mmol) and AZADO (0.50 mg, 3.34 µmol). After stirring for 1 h at rt, the reaction was quenched with sat. Na₂S₂O₃ (0.5 mL). After stirring for 30 min, the mixture was extracted with EtOAc and the combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/Hexane 1/10-1/8) gave ketone (+)-52 (8.3 mg, 6.34 μ mol, 95% over 2 steps) as a white foam.

(+)-52: $[\alpha]_{D}^{27}$ +58.9 (c 0.32, CHCl₃); IR (neat): 2947, 2360, 1725, 1590, 1469, 1353, 1247, 1172, 1083 cm⁻¹; ¹H NMR (600 MHz, $CDCl_3$) δ : 6.33 (s, 1H), 5.65 (d, J = 4.2 Hz, 1H), 5.07 (d, J = 5.4 Hz, 1H), 4.82 (s, 1H), 4.78 (s, 1H), 4.70 (s, 2H), 4.41-4.28 (m, 4H), 4.25-4.20 (m, 1H), 3.83 (dd, J = 9.6, 2.4 Hz, 1H), 3.67-3.58 (m, 3H), 3.41 (s, 3H), 3.31 (s, 3H), 3.09 (d, J = 16.2 Hz, 1H), 2.88 (dd, J = 15.0, 6.6 Hz, 1H), 2.79 (dd, J = 12.6, 4.2 Hz, 1H), 2.63 (dd, J = 15.0, 6.0 Hz, 1H), 2.30 (d, J = 6.0 Hz, 2H), 2.19 (s, 3H), 2.05–2.00 (m, 1H), 1.77 (s, 3H), 1.73–1.71 (m, 2H), 1.56 (s, 3H), 1.32 (sept, J = 7.6 Hz, 6H), 1.16-1.12 (m, 39H), 1.05 (s, 3H), 0.90-0.82 (m, 13H), 0.10-0.04 (m, 24H); ¹³C NMR (150 MHz, CDCl₃) δ: 211.8, 174.4, 163.4, 158.7, 153.9, 142.6, 141.6, 112.9, 110.4, 109.8, 94.7, 90.1, 81.1, 78.8, 68.8, 66.6, 65.9, 58.1, 57.4, 49.1, 40.8, 39.0, 38.9, 38.3, 34.1, 30.2, 29.8, 25.8, 23.1, 22.3, 19.2, 18.1, 18.1, 18.0, 17.6, 13.3, 13.2, 12.1, 9.2, -1.6, -3.3, -4.7; MS (ESI) calculated for C₆₇H₁₂₆NO₁₄Si₅ [M + H]⁺: 1308.8019, found 1308. 7990.

(-)-Irciniastatin B [(-)-2]. To a solution of ketone (+)-52 (7.0 mg, 5.35 μ mol) in DMF (0.3 mL) was added TASF (73.6 mg, 0.267 mmol) in DMF (0.1 mL), and the reaction was heated to 50 °C. After stirring for 10 h, the reaction was quenched with sat. NH₄Cl (1 mL) and the mixture was extracted with EtOAc. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated. Purification by column chromatography (CHCl₃/MeOH 1%-2%) gave (-)-irciniastatin B [(-)-2] (1.78 mg, 2.93 μ mol, 55%) as a white foam.

(-)-2: $[\alpha]_D^{27}$ -26.9 (c 0.089, MeOH) { $[\alpha]_D^{20}$ -28.7 (c 0.2, MeOH)⁹}; IR (neat): 3357, 2927, 1714, 1656, 1618, 1508, 1380, 1251, 1089 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ : 11.13 (s, 1H), 7.30 (d, *J* = 10.2 Hz, 1H), 6.27 (s, 1H), 5.17 (dd, *J* = 9.6, 6.0 Hz, 1H), 4.79

(s, 1H), 4.76 (s, 1H), 4.54 (dd, J = 8.4, 4.8 Hz, 1H), 4.41 (s, 1H), 4.18 (q, J = 6.0 Hz, 1H), 4.06 (d, J = 10.2 Hz, 1H), 3.98 (dd, J = 11.4, 1.2 Hz, 1H), 3.75–3.72 (m, 1H), 3.70 (brs, 1H), 3.60 (brs, 1H), 3.36 (s, 3H), 3.35(s, 3H), 2.95–2.88 (m, 2H), 2.64 (d, J = 7.2 Hz, 1H), 2.34 (dd, J = 15.0, 8.4 Hz, 1H), 2.12–2.10 (m, 1H), 2.07 (s, 3H), 1.89 (m, 1H), 1.83–1.80 (m, 1H), 1.73 (s, 3H), 1.63–1.60 (m, 1H), 1.14 (s, 3H), 1.10 (d, J = 7.8 Hz, 3H), 1.08 (s, 3H); ¹³C NMR (150 MHz, CDCl₃) δ : 176.4, 173.6, 170.5, 161.1, 142.0, 139.7, 113.2, 113.0, 101.6, 101.3, 81.9, 80.6, 79.6, 78.3, 73.9, 73.1, 73.1, 71.4, 57.9, 56.3, 42.7, 38.8, 37.6, 32.2, 29.7, 28.5, 23.1, 22.7, 13.6, 10.5, 9.2; MS (ESI) calculated for C₃₁H₄₆NO₁₁ [M + H]⁺: 608.3065, found 608.3057.

Toxicity. Cell survival was determined by a WST-8 assay kit. HeLa, 3Y1, MCAS, and SKOV3 cells (3×10^3 cells/well) were seeded onto 96-well plates and incubated overnight. Then, cells were treated with various concentrations of irciniastatins A and B. After 48 h incubation, 10 μ L of WST-8 reagent was added to the culture. After 2 h incubation, the absorbance at 450 nm was measured with an iMark microplate reader (BioRad Laboratories, Inc.). The number of living cells (% control) was calculated with the following formula: (each absorbance – absorbance of blank well)/(absorbance of 0 μ M well – absorbance of blank well) × 100.

Protein Synthesis Assay. HeLa and 3Y1 cells $(1.25 \times 10^4 \text{ cells}/\text{ well})$ were seeded onto a 24-well plate in Dulbecco's modified MEM (DMEM) supplemented with 10% FCS and incubated overnight. Then, cells were treated with various concentrations of irciniastatins A and B for 2 h, followed by [methyl-³H] methionine (finally 3.7 kBq/mL) addition. After 2 h incubation, the cells were washed with PBS once, and ice-cold 5% TCA was added. Acid-insoluble fractions were solubilized by 0.25 M NaOH, and the radioactivity was determined using an LS 6500 liquid scintillation counter (Beckman Coulter Inc., Brea, CA).

Chemical Footprinting Assay.³⁰ 80S ribosomes were purified from yeast strain BY4741 by two cycled ultracentrifugation through sucrose cushions. 60 pmol of ribosomes, with a total volume of 80 μ L, was treated with each compound for 5 min at 25 °C. After drug treatment, 20 μ L of DMS was added to final 20 mM or 90 mM, and reacted for 5 min at 37 °C. The reaction was guenched, and rRNA was extracted by Phe/Chl extraction. 2.0 μg of rRNA treated with each drug was mixed with ³²P labeled primer (TGTCGCTAT-GAACGCTTGACTG, annealing at 2853-2832 base of yeast 25S rRNA), and annealed by heating to 60 °C for 20 min and cooling on ice for 5 min. Then, primer extension was performed using AMV reverse transcriptase for 45 min at 43 °C. The reaction was stopped by adding 2 × loading buffer (98% formamide, 10 mM EDTA, 0.1% xylene cyanol, 0.1% bromophenol blue) and heating to 90 °C for 10 min. These samples were loaded onto 8% polyacrylamide sequencing gel, and separated by 1600 V electrophoresis for 2 h. The gel was dried, and ³²P radioactivity was measured by Typhoon 8600.

ASSOCIATED CONTENT

S Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.joc.5b02256.

¹H and ¹³C spectral data of compounds and HPLC chromatograms (PDF)

AUTHOR INFORMATION

Corresponding Authors

*E-mail: usui.takeo.kb@u.tsukuba.ac.jp (T.U.).

*E-mail: y-iwabuchi@m.tohoku.ac.jp (Y.I.).

Notes

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REFERENCES

(1) Pettit, G. R.; Xu, J.-P.; Chapuis, J.-C.; Pettit, R. K.; Tackett, L. P.; Doubek, D. L.; Hooper, J. N. A.; Schmidt, J. M. *J. Med. Chem.* **2004**, 47, 1149.

(2) Cichewicz, R. H.; Valeriote, F. A.; Crews, P. Org. Lett. 2004, 6, 1951.

(3) Jiang, X.; García-Fortanet, J.; De Brabander, J. K. J. Am. Chem. Soc. 2005, 127, 11254.

(4) (a) Huang, X.; Shao, N.; Palani, A.; Aslanian, R.; Buevich, A. Org. Lett. 2007, 9, 2597. (b) Smith, A. B., III; Jurica, J. A.; Walsh, S. P. Org. Lett. 2008, 10, 5625. (c) Crimmins, M. T.; Stevens, J. M.; Schaaf, G. M. Org. Lett. 2009, 11, 3990. (d) Watanabe, T.; Imaizumi, T.; Chinen, T.; Nagumo, Y.; Shibuya, M.; Usui, T.; Kanoh, N.; Iwabuchi, Y. Org. Lett. 2010, 12, 1040. (e) Wan, S.; Wu, F.; Rech, J. C.; Green, M. E.; Balachandran, R.; Horne, W. S.; Day, B. W.; Floreancig, P. E. J. Am. Chem. Soc. 2011, 133, 16668. (f) Byeon, S. R.; Park, H.; Kim, H.; Hong, J. Org. Lett. 2011, 13, 5816. (g) Feng, Y.; Jiang, X.; De Brabander, J. K. J. Am. Chem. Soc. 2012, 134, 17083.

(5) (a) Shangguan, N.; Kiren, S.; Williams, L. J. Org. Lett. 2007, 9, 1093. (b) Bielitza, M.; Pietruszka, J. Chem. - Eur. J. 2013, 19, 8300.

(6) (a) Pal, A.; Peng, Z.; Schuber, P. T., Jr.; Prasad, B. A. B.; Bornmann, W. G. *Tetrahedron Lett.* **2013**, *54*, 5555 and other references cited in ref 8.

(7) (a) Jiang, X.; Williams, N.; De Brabander, J. K. Org. Lett. 2007, 9, 227. (b) Huang, X.; Shao, N.; Huryk, R.; Palani, A.; Aslanian, R.; Seidel-Dugan, C. Org. Lett. 2009, 11, 867. (c) Shao, N.; Huang, X.; Palani, A.; Aslanian, R.; Buevich, A.; Piwinski, J.; Huryk, R.; Seidel-Dugan, C. Synthesis 2009, 2009, 2855. (d) Wu, C.-Y.; Feng, Y.; Cardenas, E. R.; Williams, N.; Floreancig, P. E.; De Brabander, J. K.; Roth, M. G. J. Am. Chem. Soc. 2012, 134, 18998.

(8) For the review of results in the past decade, see: Bielitza, M.; Pietruszka, J. Angew. Chem., Int. Ed. 2013, 52, 10960.

(9) (a) An, C.; Hoye, A. T.; Smith, A. B., III Org. Lett. 2012, 14, 4350. (b) An, C.; Jurica, J. A.; Walsh, S. P.; Hoye, A. T.; Smith, A. B., III J. Org. Chem. 2013, 78, 4278.

(10) During the SAR study, it was found that the C11-deoxy analogue possesses more potent cytotoxicity than (+)-1 (see refs 7b and 7c).

(11) (a) Katsuki, T.; Sharpless, K. B. J. Am. Chem. Soc. **1980**, 102, 5974. (b) Hanson, R. M.; Sharpless, K. B. J. Org. Chem. **1986**, 51, 1922.

(12) Alegret, C.; Santacana, F.; Riera, A. J. Org. Chem. 2007, 72, 7688.
(13) (a) Caron, M.; Sharpless, K. B. J. Org. Chem. 1985, 50, 1557.

(b) Chong, J. M.; Sharpless, K. B. J. Org. Chem. 1985, 50, 1560.
(c) Behrens, C. H.; Ko, S. Y.; Sharpless, K. B.; Walker, F. J. J. Org. Chem. 1985, 50, 5687. (d) Behrens, B. H.; Sharpless, K. B. J. Org. Chem. 1985, 50, 5696.

(14) For the screening of reaction conditions and application to other substrates and nucleophiles, see: Uesugi, S.; Watanabe, T.; Imaizumi, T.; Shibuya, M.; Kanoh, N.; Iwabuchi, Y. *Org. Lett.* **2014**, *16*, 4408.

(15) Shibuya, M.; Tomizawa, M.; Suzuki, I.; Iwabuchi, Y. J. Am. Chem. Soc. 2006, 128, 8412.

(16) Lavallée, P.; Ruel, R.; Grenier, L.; Bissonnette, M. Tetrahedron Lett. **1986**, 27, 679.

(17) Yamaguchi, M.; Hirao, I. Tetrahedron Lett. 1983, 24, 391.

(18) De Brabander, J. K.; Vandewalle, M. *Synthesis* **1994**, *1994*, 855. (19) The stereochemistry of (+)-**23** was determined by NOE correlations.

(20) Shibuya, M.; Sato, T.; Tomizawa, M.; Iwabuchi, Y. Chem. Commun. 2009, 1739.

(21) Hamana, H.; Sasakura, K.; Sugasawa, T. Chem. Lett. 1984, 13, 1729.

(22) Evans, D. A.; Calter, M. A. Tetrahedron Lett. 1993, 34, 6871.

(23) Although we tried TBS-, Bz-, and SEM-groups for protection, only the SEM-protected side chain (-)-6 could be reacted. In the case of TBS- and Bz-groups, the expected reaction did not occur and the starting material was recovered.

(24) The structure of (+)-40 was determined by a detailed analysis of the HMBC and COSY spectra; see the Supporting Information.

(25) Keck, G. E.; Truong, A. P. Org. Lett. 2005, 7, 2153.

(26) Doi, R.; Shibuya, M.; Murayama, T.; Yamamoto, Y.; Iwabuchi, Y. J. Org. Chem. **2015**, 80, 401.

(27) In NOESY spectra, the correlation between the C8-proton (δ 5.17) and the C13-proton (δ 3.98) was observed.

(28) Chinen, T.; Nagumo, Y.; Watanabe, T.; Imaizumi, T.; Shibuya, M.; Kataoka, T.; Kanoh, N.; Iwabuchi, Y.; Usui, T. *Toxicol. Lett.* **2010**, 199, 341.

(29) Dang, Y.; Schneider-Poetsch, T.; Eyler, D. E.; Jewett, J. C.; Bhat, S.; Rawal, V. H.; Green, R.; Liu, J. O. *RNA* **2011**, *17*, 1578.

(30) Schneider-Poetsch, T.; Ju, J.; Eyler, D. E.; Dang, Y.; Bhat, S.; Merrick, W. C.; Green, R.; Shen, B.; Liu, J. O. *Nat. Chem. Biol.* **2010**, *6*, 209.